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**Kinetic study of the supercritical CO₂ extraction of
different plants from *Lamiaceae* family**

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Running title: CO₂ extraction of *Lamiaceae* plants.

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23 **Abstract**

24 The supercritical CO₂ extraction of four different plants from *Lamiaceae* family,
25 namely oregano (*Origanum vulgare*), thyme (*Thymus zygis*), sage (*Salvia officinalis*)
26 and rosemary (*Rosmarinus officinalis*) was carried out in an experimental pilot-plant
27 comprising an extraction cell of two liters capacity. 600 g of leaves of each plant
28 material, with the same pre-treatment, were extracted at the same pressure and
29 temperature (30 MPa and 313 K) and using 2.4 kg/h of CO₂. Further, the same
30 fractionation procedure in a two on-line decompressing separators at, respectively, 10
31 MPa and 0.1 MPa was employed. In this way, a thoughtful comparison of the
32 extraction kinetic was established and discussed, in terms of the extraction yields
33 attained in the separators, the variation of the essential oil composition with time and
34 the content of key bioactive substances identified in the different fractions.

39 **Keywords:** supercritical extraction; carbon dioxide; oregano; sage; rosemary; thyme.

42 **1. Introduction**

43 In the European market there are a lot of products derived from natural plants,
44 commonly recognized with biological properties, such as antioxidant, antiseptic,
45 diuretic, stimulating the central nervous system, sedative, expectorant, digestive, etc.

46 Some of these plants have been used in traditional medicine since ancient times and
47 are available on market as infusions, tablets and/or extracts.

48 Natural sources of bioactive substances, as well as new industrial approaches to
49 extract and isolate these substances from raw materials, are gaining much attention in
50 the food and pharmaceutical research field. Indeed, among innovative process
51 technologies, supercritical CO₂ (SC-CO₂) extraction and fractionation is the most
52 widely studied application. The production of supercritical plant extracts has received
53 increasing interest in recent decades [1-3] and has brought a wide variety of products
54 that are being intensively investigated due to their favorable effects on diversity
55 human diseases. Different authors compared supercritical extracts with those obtained
56 using liquid solvents (ethanol and hexane) or hydrodistillation, and described superior
57 quality (better functional activity) of the supercritical extracts [4-5].

58 Among the different vegetable raw materials considered, several plants from the
59 *Lamiaceae* family were subject of intensive study. In general, the essential oils of
60 these plants are recognized to contain the substances for which the plant is used in the
61 pharmaceutical, food or fragrance industries. Essential oils represent a small fraction
62 of the plant composition; the main compounds are terpenes and sesquiterpenes, and
63 several oxygenated derivatives compounds (alcohols, aldehydes, ketones, acids,
64 phenols, ethers, esters, etc.) all of them responsible for the characteristic plant odor
65 and flavor [2].

66 Particularly, *Origanum vulgare L.* is an herbaceous plant native of the Mediterranean

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67 regions, used as a medicinal plant with healthy properties like its powerful anti-
68 bacterial and anti-fungal properties [6, 7]. The responsible of these activities in
69 oregano is the volatile oil, which contains thymol and carvacrol as the primary
70 components [8]. In these compounds, Puertas-Mejia et al. [9] also found some
71 antioxidant activity.

72 The supercritical extraction and fractionation of oregano has been studied and
73 reported in the literature [10 - 12]. Moderate conditions (solvent densities between
74 300 and 500 kg/m³) were found to be sufficient for an efficient extraction of volatile
75 oil compounds. Although higher pressures increase the rate of extraction and yield of
76 the essential oil fraction, also significant amounts of waxes were co-extracted and,
77 consequently, the essential oil content in the extract decreased [12].

78 Thymol and carvacrol were also found in the essential oil of another *Lamiaceae* plant,
79 namely *Thymus*. The variety most studied is, indeed, *Thymus vulgaris* [13-14]. Yet,
80 particularly attention is focused on *Thymus zygis*, a thyme variety widespread over
81 Portugal and Spain, which extract has proved to be useful for food flavoring [15] and
82 in the pharmaceutical [16-17] and cosmetic industries [18]. Moldao-Martins et al. [19]
83 studied the supercritical extraction of *Thymus zygis* at different temperatures (300-323
84 K) and pressures (8-20 MPa) and reported a comprehensive comparison of the
85 extracts produced with those obtained from steam distillation.

86 Other *Lamiaceae* plants being intensively studied are the "*Officinalis*" ones (from
87 Latin meaning medicinal). Sage (*Salvia officinalis* L.) is a popular kitchen herb and
88 has been used in a variety of food preparations since ancient times, and has a
89 historical reputation for promotion of health and treatment of diseases [20]. Modern
90 day research has shown that sage essential oil can improve the memory and has
91 shown promise in the treatment of Alzheimer's disease [21]. In the past few decades

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92 however, sage has been the subject of an intensive study for its phenolic antioxidant
93 components [22-24]. Supercritical extraction of sage demonstrated that when sage
94 leaves are ground in fine particles, the essential oil is easily accessible to the SC-CO₂
95 solvent (9-13 MPa and 25-50 °C) and the extraction is controlled by phase
96 equilibrium [25]. That is, large part of the total essential oil contained in the plant
97 matrix is dissolved almost immediately in SC-CO₂. To extract high molecular and
98 polar compounds from sage, CO₂ with an ethanol-water mixture as co-solvent was
99 employed; antioxidant substances such as rosmarinic acid and carnosic compounds
100 were extracted, achieving a recovery of 55 % and 75 % respectively [26].

101 The supercritical extraction of rosemary (*Rosmarinus officinalis L.*), which has been
102 recognized as one of the plants with large antioxidant activity, also produced extracts
103 with large concentrations of phenolic antioxidants. Main substances associated with
104 the antioxidant activity of rosemary extract are the phenolic diterpenes such as
105 carnosol, rosmanol, carnosic acid, methyl carnosate, and phenolic acids such as the
106 rosmarinic and caffeic acids [27-31]. Among the large number of papers related with
107 the supercritical extraction and fractionation of rosemary and its effect on the
108 antioxidant activity of the extracts, the authors have recently presented two new
109 contributions [32, 33].

110 Indeed, numerous variables have singular effect on the supercritical extraction yield
111 and on the composition and quality of extracts. Process conditions, such as extraction
112 pressure and temperature, type and amount of cosolvent, extraction time,
113 fractionation, raw material pre-treatment, plant location and harvesting time, greatly
114 affect not only yield but also composition of the extracted material. The different
115 process conditions applied, together with the variety of equipment and process scale
116 employed, complicate the comparison of the competence of supercritical CO₂

117 technology in the extraction of bioactive compounds from plant material.

118 In this paper we carried out the extraction of four *Lamiaceae* plant varieties, namely
119 oregano, thyme, sage and rosemary, using the same procedure for the preparation of
120 the raw materials (plant leaves), employing the same experimental pilot-plant device
121 and the same extraction conditions and procedure. Then, the kinetic behavior of the
122 extractions, considering both yield and composition of the fractions obtained, was
123 evaluated and compared.

125 **2. Materials and methods**

126 **2.1 Chemicals**

127 Carnosic acid ($\geq 96\%$) were purchased from Alexis Biochemical (Madrid, Spain).
128 Thymol (99.5%), Camphor ($>97\%$) and Linalool ($>97\%$) were purchased from
129 SIGMA-ALDRICH (Madrid, Spain), whereas 1,8 cineole (98%) and Borneol ($>99\%$)
130 were purchased from FLUKA (Madrid, Spain). Ethanol, acetonitrile and phosphoric
131 acid were all HPLC grade from Lab Scan (Dublin, Ireland).

132 **2.2 Preparation of plant leaves**

133 Plant material consisted of dried leaves obtained from an herbalist's producer
134 (Murcia, Spain). A kitchen-type knife mill was employed to carry out grinding of the
135 leaves. The mill was adapted so as to break up the raw material under cryogenic
136 conditions (using carbon dioxide). The particle size distribution was determined with
137 a vibratory sieve shaker. Sieves were selected in order to have high yield in the
138 grinding process ($>85\%$). Particle size obtained was in the range of 500 to 1000 μm .
139 The samples were stored at -20°C until use.

140 **2.3 Supercritical extraction method**

141 Extractions were carried out in a pilot-plant scale supercritical fluid extractor (Thar

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142 Technology, Pittsburgh, PA, USA, model SF2000) comprising a 2 L cylinder
143 extraction cell and two different separators (S1 and S2), each of 0.5 L capacity, with
144 independent control of temperature and pressure. The extraction vessel has a
145 height/diameter ratio of 5.5 (0.42 m height, 0.076 m internal diameter). A detail
146 explanation of the experimental device can be found elsewhere [34].

147 For each experiment, the cell was filled with 0.6 kg of plant raw material. The
148 extractions were performed at a pressure constant of 30 MPa. Fractionation of the
149 extract was accomplished maintaining S1 at 10 MPa and S2 at ambient pressure (0.1
150 MPa). Extraction and fractionation pressure was set to be 313 K in all experimental
151 assays. Further, CO₂ flow rate was set to 2.4 kg/h in all experiments (CO₂/plant = 20
152 kg/kg). Samples were collected from both separators at intervals of 1.5 h during 4.5 h.
153 The solid fractions obtained in S1 and S2 were recuperated and placed in vials. In
154 order to ensure an accurate determination of extraction yield with time, separators
155 were washed with ethanol and the residual material recovered in each case was mixed
156 with the corresponding solid fraction. Ethanol was eliminated by evaporation and
157 then, homogeneous solid samples were obtained and kept under N₂ at -20°C in the
158 dark until analysis.

159 **2.4 HPLC analysis**

160 In order to quantify the carnosic acid content in the rosemary extracts, samples were
161 analyzed employing a HPLC (Varian Pro-star) equipped with a Nova Pack C18
162 column (Waters) of 15 mm × 4.6 mm and 3.5 μm particle size. The mobile phase
163 consisted of acetonitrile (solvent A) and 0.1% of phosphoric acid in water (solvent B)
164 applying the following gradient: 0–8 min, 23% A and 8–20 min, 75% A. This last
165 composition was kept until the end of the chromatogram and initial conditions were
166 gained in 5 min. Total time analysis was 40 minutes. The flow rate was constant at 0.7

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167 mL/min. Injection volume was 20 μ L and the detection was accomplished by using a
168 diode array detection system Varian storing the signal at a wavelength of 230, 280
169 and 350 nm.

170 **2.5 GC-MS analysis**

171 Oregano, sage and thyme extracts were analyzed by GC-MS in order to determine the
172 essential oil composition of the different fractions collected. In the case of oregano
173 and sage, a GC-2010 (Shimadzu, Japan) was employed, comprising a split/splitless
174 injector, electronic pressure control, AOC-20i auto injector, GCMS-QP2010 Plus
175 mass spectrometer detector, and GC-MS Solution software. The column used was a
176 ZB-5 (Zebron) capillary column, 30 m x 0.25 mm I.D. and 0.25 μ m phase thickness.
177 For thyme extracts, a 7890A System (Agilent Technologies, U.S.A.) was employed,
178 comprising a split/splitless injector, electronic pressure control, G4513A auto injector,
179 a 5975C triple-Axis mass spectrometer detector, and GC-MS Solution software. The
180 column used was an Agilent 19091S-433 capillary column, 30 m x 0.25 mm I.D. and
181 0.25 μ m phase thickness. For all the analysis, the chromatographic method was as
182 follows: oven temperature programming was 60 $^{\circ}$ C isothermal for 4 min then
183 increased to 106 $^{\circ}$ C at 2.5 $^{\circ}$ C/min and from 106 $^{\circ}$ C to 130 $^{\circ}$ C at 1 $^{\circ}$ C/min and finally
184 from 130 $^{\circ}$ C to 250 $^{\circ}$ C at 20 $^{\circ}$ C/min, this temperature was kept constant for 10.2 min.
185 Sample injections (1 μ L) were performed in split mode (1:20). Helium, 99.996% was
186 used as a carrier gas at a flow of 1 mL/min with an inlet pressure of 57.5 KPa. Injector
187 temperature was of 250 $^{\circ}$ C and MS ion source and interface temperatures were 230 $^{\circ}$ C
188 and 280 $^{\circ}$ C, respectively. The mass spectrometer was used in TIC mode, and samples
189 were scanned from 40 to 500 amu. Thymol, borneol, camphor, 1,8 cineole and
190 linalool were identified by comparison with standard mass spectra, obtained in the
191 same conditions and compared with the mass spectra from library Wiley 229. Rests of

192 the compounds were identified by comparison with the mass spectra from Wiley 229
193 library. A calibration curve was employed to quantify thymol, camphor and carnosic
194 acid content.

195

196 **3. Results and discussion**

197 Table 1 show the amounts of material recovered in each separator (S1 and S2) during
198 each interval of time (first interval: 0-1.5 h; second interval: 1.5- 3 h; and third
199 interval: 3-4.5 h) for the four plants extracted. Figure 1 show a comparison between
200 the global yields (S1 + S2) obtained for the different raw materials as a function of
201 extraction time. As can be deduced from the figure, salvia and oregano were
202 completely extracted, with an estimated optimal extraction time of 1.76 h (see Figure
203 1). But in the case of rosemary and thyme, none of these plant materials were
204 completely exhausted during the 4.5 h of extraction. Moreover, very similar kinetic
205 behavior resulted for salvia and oregano, so as for thyme and rosemary. Considering
206 the first period of time (t_1 : 0 - 1.5 h) it was estimated a removal velocity of around
207 0.004 g extract / g CO₂ in the case of salvia and oregano, and almost half of this value
208 in the case of rosemary and thyme.

209 With respect to the fractionation of the extracted material, the performance is quite
210 different considering the diverse plants studied (see Table 1). In the case of oregano,
211 the amount of material recovered in S2 is almost half the amount recovered in S1. Just
212 the opposite behavior is observed for sage and thyme, while in the case of rosemary
213 extraction similar amounts of extract were recovered in both S1 and S2.

214 Despite the distinct fractionation behavior observed that definitely should be
215 attributed to the different substances that compose the extracts (extraction and
216 fractionation conditions were kept exactly the same), the essential oil compounds

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217 were selectively recovered in S2 separator for the four plant materials studied.

218 Tables 2, 3 and 4 present the essential oil compounds identified, respectively, in

219 oregano, sage and thyme extracts, according to the GC-MS analysis. These tables

220 provide the total area determined for each compound (Tables 2a, 3a and 4a) and the

221 essential oil composition in terms of the percentage of area of each identified

222 substance (Tables 2b, 3b and 4b). As can be deduced from the tables, the main

223 compounds identified in oregano were thymol, sabinene hydrate and carvacrol. In the

224 case of sage extractions, the main substances detected were camphor and cineole,

225 following by borneol and sabinyl and linalyl acetates. Finally, for thyme extracts the

226 main compounds identified were thymol and N-II (a non-identified compound with a

227 retention time of 49.09 min) following by carvacrol and borneol.

228 As mentioned before, a concentration of the volatile oil compounds is selectively

229 produced in S2 for all plants studied. The ratio between the total area quantified in S2

230 and the total area quantified in S1 ($S2/S1$) is, respectively, 9.7, 3.4 and 14.2 for

231 oregano, sage and thyme (see Tables 2 to 4). This means that 90.6, 77.6 and 93.4 % of

232 the volatile oil compounds identified, respectively, in oregano, sage and thyme were

233 recovered in S2 separator. This selectively recovery in S2 of the essential oil

234 compounds come to an agreement with the higher extractions yields obtained in this

235 separator in the case of sage and thyme. But it is clear that in oregano extraction, high

236 amounts of substances different from the volatile oil compounds are extracted and

237 precipitated in S1 separator (i.e. the high yield obtained in S1 oregano extraction is

238 not due to the volatile oil removal).

239 Figures 2 and 3 show the variation with time of the quantified areas obtained for the

240 main compounds identified in the S1 samples (Figure 2) and in the S2 samples

241 (Figure 3) of oregano, sage and thyme. In general, as expected, the concentration of

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242 these compounds decrease with time both in S1 and S2 samples. Further, a noticeable
243 reduction in the extraction of these compounds is observed in the case of oregano and
244 sage, what agree with the fact that oregano and sage leaves are almost exhausted
245 during the first interval of extraction (0 - 1.5 h). But in the case of thyme extracts, the
246 decrease in the essential oil compounds extraction is much less pronounced, what
247 approves the delayed kinetic behavior observed in thyme leaves.

248 The concentrations (% weight) of some key components with recognized biological
249 activity were also determined and are given in Table 5: thymol in oregano and thyme
250 extracts, camphor in sage, and carnosic acid in rosemary. As expected, the % weight
251 of the monoterpene compounds (thymol and camphor), which are main constituents of
252 the volatile oil fractions, decrease with extraction time. But the concentration of
253 carnosic acid in the rosemary fractions recovered, increase with extraction time.
254 Further, 72.4 % of the total antioxidant carnosic acid extracted from rosemary was
255 selectively recovered in the first separator.

256 Decreasing percentages of lighter compounds (terpenes and oxygenated terpenes)
257 were found as extraction time increase, while higher-molecular-weight compounds,
258 such as a phenolic diterpene, showed a continuous percentage increase at increasing
259 extraction times, as observed by Reverchon et al. [35]. As sake of comparison, it was
260 calculated that 97.6 % of the mass of camphor extracted from sage was precipitated in
261 S1 and S2 separators during the first interval of time (t1). Also high recoveries and
262 very similar values were obtained for the recovery of thymol during t1: 82.6 and 80.4
263 %, respectively, in the oregano and thyme extraction. All these values are
264 significantly higher than the recovery obtained for the carnosic acid extracted from
265 rosemary during t1 (41.4 %). Furthermore, these values agree with the order reported
266 in the literature [36, 37] for the solubility of these substances in supercritical CO₂

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267 (camphor > thymol >> carnosic acid).

268

269 **Conclusion**

270 The supercritical extraction of *Lamiaceae* plants, namely oregano, sage, thyme and
271 rosemary, was carried out under identical conditions of raw material pre-treatment,
272 apparent density in the extraction cell, extraction pressure and temperature and
273 fractionation procedure. Oregano and sage were much more rapid exhausted than
274 thyme and rosemary, presenting very similar kinetic behavior in terms of extraction
275 yield. The fractionation of the extract indicated that sage and thyme contains larger
276 amounts of high volatile or high CO₂ soluble substances than oregano or rosemary,
277 since for sage and thyme the yield obtained in S2 was almost double the yield
278 obtained in S1. Thymol, a monoterpene phenol which is one of the main components
279 of oregano and thyme plants, was highly extracted despite the plant variety: 82.6 and
280 80.4 % of the total amounts of thymol present in, respectively, oregano and thyme
281 extracts were recovered during the first interval of extraction. On the other side,
282 carnosic acid was only 41.4 % recovered from rosemary in this extraction period.
283 Thus, the weight content of lighter compounds (thymol and camphor) were found to
284 decrease with extraction time, while the weight content of higher molecular weight
285 and less soluble substance (carnosic acid) showed a continuous increase at increasing
286 extraction times.

287

288 **Acknowledges**

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401 **Table 1.** Mass (g) of material recovered in each separator cell (S1 and S2) as a
 402 function of time in the extraction of oregano, sage, thyme and rosemary at 30 MPa
 403 and 313 K.

404

	time (h)	S1 (g)	S2 (g)
oregano	1.5	15.513	7.211
	3.0	3.346	2.006
	4.5	0.203	0.325
	global yield	19.062	9.542
sage	1.5	6.794	16.359
	3.0	1.261	2.708
	4.5	0.269	0.311
	global yield	8.324	19.378
thyme	1.5	3.720	6.800
	3.0	1.220	1.930
	4.5	0.510	1.490
	global yield	5.45	10.22
rosemary	1.5	6.287	5.599
	3.0	2.083	2.750
	4.5	2.220	2.135
	global yield	10.59	10.484

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408 **Table 2.** Chromatographic GC-MS areas of the essential oil compounds identified in
 409 the oregano extracts as a function of time. t1, t2 and t3 correspond to the three
 410 intervals of time studied.

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412 (a) *Absolute areas*

Retention time	Compound	S1			S2		
		t1	t2	t3	t1	t2	t3
13.35	Limonene	-	-	-	145257	123798	107069
14.94	γ -Terpinene	59200	-	-	45281	103523	-
15.38	cis-Sabinene hydrate	307051	-	-	2080399	920461	433226
17.18	trans-Sabinene hydrate	6968774	617088	965065	55412749	21087412	9946755
17.36	Linalool	184257	13823	39405	1699025	800394	425076
21.75	Terpineol	439285	34796	46977	3975567	1355249	633420
22.54	α -terpineol	505669	43278	60984	4644332	1453346	683040
25.68	Thymyl methyl ether	104106	-	-	1180444	463215	207421
26.20	Sabinene hydrate acetate	119882	-	-	1870980	452084	186150
26.43	Linalyl acetate	165154	-	-	2677209	779175	343339
28.70	Thymol	6378950	546737	898247	43532669	15690062	7463979
29.28	Carvacrol	2487757	189595	286686	16611808	5399309	2307413
37.85	E-caryophyllene	231111	-	-	3481108	938718	457200
Total area (t1+t2+t3)		21693877			209741538		

413

414 (b) *Percentage area*

Retention time	Compound	S1			S2		
		t1	t2	t3	t1	t2	t3
13.35	Limonene	-	-	-	0.11	0.25	0.46
14.94	γ -Terpinene	0.33	-	-	0.03	0.21	-
15.38	cis-Sabinene hydrate	1.71	-	-	1.51	1.86	1.87
17.18	trans-Sabinene hydrate	38.82	42.70	42.01	40.34	42.54	42.88
17.36	Linalool	1.03	0.96	1.72	1.24	1.61	1.83
21.75	Terpineol	2.45	2.41	2.04	2.89	2.73	2.73
22.54	α -terpineol	2.82	2.99	2.65	3.38	2.93	2.94
25.68	Thymyl methyl ether	0.58	-	-	0.86	0.93	0.89
26.20	Sabinene hydrate acetate	0.67	-	-	1.36	0.91	0.80
26.43	Linalyl acetate	0.92	-	-	1.95	1.57	1.48
28.70	Thymol	35.53	37.83	39.10	31.69	31.65	32.18
29.28	Carvacrol	13.86	13.12	12.48	12.09	10.89	9.95
37.85	E-caryophyllene	1.29	-	-	2.53	1.89	1.97

415

416 **Table 3.** Chromatographic GC-MS areas of the essential oil compounds identified in
 417 the sage extracts as a function of time. t1, t2 and t3 correspond to the three intervals of
 418 time studied.

419 (a) *Absolute areas*

Retention time	Compound	S1			S2		
		t1	t2	t3	t1	t2	t3
13.30	1.8 cineole	499567	104213	47435	2241357	159084	41305
15.38	Cis sabinene hydrate	40740	-	-	118065	-	-
17.18	Trans Sabinene hydrate	14122	-	-	65411	38836	105568
17.36	Linalool	62288	-	-	175147	-	-
19.60	Cis sabinol	78268	-	-	310751	54752	29098
19.75	Camphor	1516240	444963	200370	5639369	679951	297560
21.05	Borneol	241000	78901	47798	954417	192116	120853
21.75	Terpineol	-	-	-	83930	-	-
22.54	α -terpineol	48370	-	-	183878	43913	29941
26.32	Geraniol	51713	-	-	151456	52063	17125
26.43	Linalyl acetate	184062	49984	-	626072	71727	25647
28.17	Endobornyl acetate	117396	36275	-	350615	55674	15919
28.68	Sabinyl acetate	179528	57611	43841	633299	156566	231019
32.58	α -terpinenyl	117363	47660	-	428938	59980	-
37.85	E-caryophyllene	80540	-	-	259271	44442	-
40.62	α -humulene	54515	-	-	186375	-	-
43.03	Geranyl propionate	66497	-	-	169758	-	-
51.18	Spathulenol	57021	-	-	147270	42472	-
51.47	Caryophyllene oxide	-	-	-	107806	-	-
52.05	Viridiflorol	79729	-	-	259112	82749	52411
Total area (t1+t2+t3)		4556509			15793068		

420 (b) *Percentage areas*

Retention time	Compound	S1			S2		
		t1	t2	t3	t1	t2	t3
13.30	1.8 cineole	14.32	13.50	13.97	17.12	9.17	4.27
15.38	Cis sabinene hydrate	1.17	-	-	0.90	-	-
17.18	Trans Sabinene hydrate	0.40	-	-	0.50	2.24	10.92
17.36	Linalool	1.79	-	-	1.34	-	-
19.60	Cis sabinol	2.24	-	-	2.37	3.16	3.01
19.75	Camphor	43.46	57.64	59.03	43.07	39.21	30.79
21.05	Borneol	6.91	10.22	14.08	7.29	11.08	12.50
21.75	Terpineol	-	-	-	0.64	-	-
22.54	α -terpineol	1.39	-	-	1.40	2.53	3.10
26.32	Geraniol	1.48	-	-	1.16	3.00	1.77
26.43	Linalyl acetate	5.28	6.48	-	4.78	4.14	2.65
28.17	Endobornyl acetate	3.36	4.70	-	2.68	3.21	1.65
28.68	Sabinyl acetate	5.15	7.46	12.92	4.84	9.03	23.90
32.58	α -terpinenyl	3.36	-	0.00	3.28	3.46	-
37.85	E-caryophyllene	2.31	-	-	1.98	2.56	-
40.62	α -humulene	1.56	-	-	1.42	-	-
43.03	Geranyl propionate	1.91	-	-	1.30	-	-
51.18	Spathulenol	1.63	-	-	1.12	2.45	-
51.47	Caryophyllene oxide	-	-	-	0.82	-	-
52.05	Viridiflorol	2.29	-	-	1.98	4.77	5.42

422 **Table 4.** Chromatographic GC-MS areas of the essential oil compounds identified in
 423 the thyme extracts as a function of time. t1, t2 and t3 correspond to the three intervals
 424 of time studied.

426 (a) *Absolute areas*

Retention n time	Compound	S1			S2		
		t1	t2	t3	t1	t2	t3
16.45	P-Cymene	30852	-	-	1751398	99903	37246
16.90	1,8 cineole	-	-	-	254488	56062	56632
19.47	Sabinene	-	-	-	197383	74813	58804
21.17	Linalool	53929	-	-	2129639	820577	471910
21.50	Trans-Sabinene Hidrate	-	-	-	174697	117889	193211
24.97	Camphor	-	-	-	537949	240541	258267
26.20	Borneol	68169	49514	49361	1871812	1087149	677940
26.52	α -Terpineol	-	-	-	209063	96019	58863
32.36	Camphene	-	-	-	237107	187021	150831
34.00	N-I	-	-	-	307110	218910	
35.00	Thymol	1335059	987930	999037	20822212	12928508	8587835
35.61	Carvacrol	89066	67342	60093	1753178	990554	638324
45.04	E-Caryophyllene	-	-	-	456217	166961	108598
49.09	N-II	249940	293465	293939	2289516	2538140	1597470
Total area (t1+t2+t3)		4627696			65510747		

427

428 (b) *Percentage areas*

Retention time	Compound	S1			S2		
		t1	t2	t3	t1	t2	t3
16.45	P-Cymene	1.69	-	-	5.31	0.51	0.29
16.90	1,8 cineole	-	-	-	0.77	0.29	0.44
19.47	Sabinene	-	-	-	0.60	0.38	0.46
21.17	Linalool	2.95	-	-	6.46	4.18	3.66
21.50	Trans-Sabinene Hidrate	-	-	-	0.53	0.60	1.50
24.97	Camphor	-	-	-	1.63	1.23	2.00
26.20	Borneol	3.73	3.54	3.52	5.67	5.54	5.26
26.52	α -Terpineol	-	-	-	0.63	0.49	0.46
32.36	Camphene	-	-	-	0.72	0.95	1.17
34.00	N-I	-	-	-	0.93	1.12	
35.00	Thymol	73.07	70.65	71.24	63.11	65.88	66.59
35.61	Carvacrol	4.87	4.82	4.28	5.31	5.05	4.95
45.04	E-Caryophyllene	-	-	-	1.38	0.85	0.84
49.09	N-II	13.68	20.99	20.96	6.94	12.93	12.39

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430 **Table 5.** Concentration (% weight) of bioactive compounds identified in oregano,
 431 sage, thyme and rosemary extracts. t1, t2 and t3 correspond to the three intervals of
 432 time studied.

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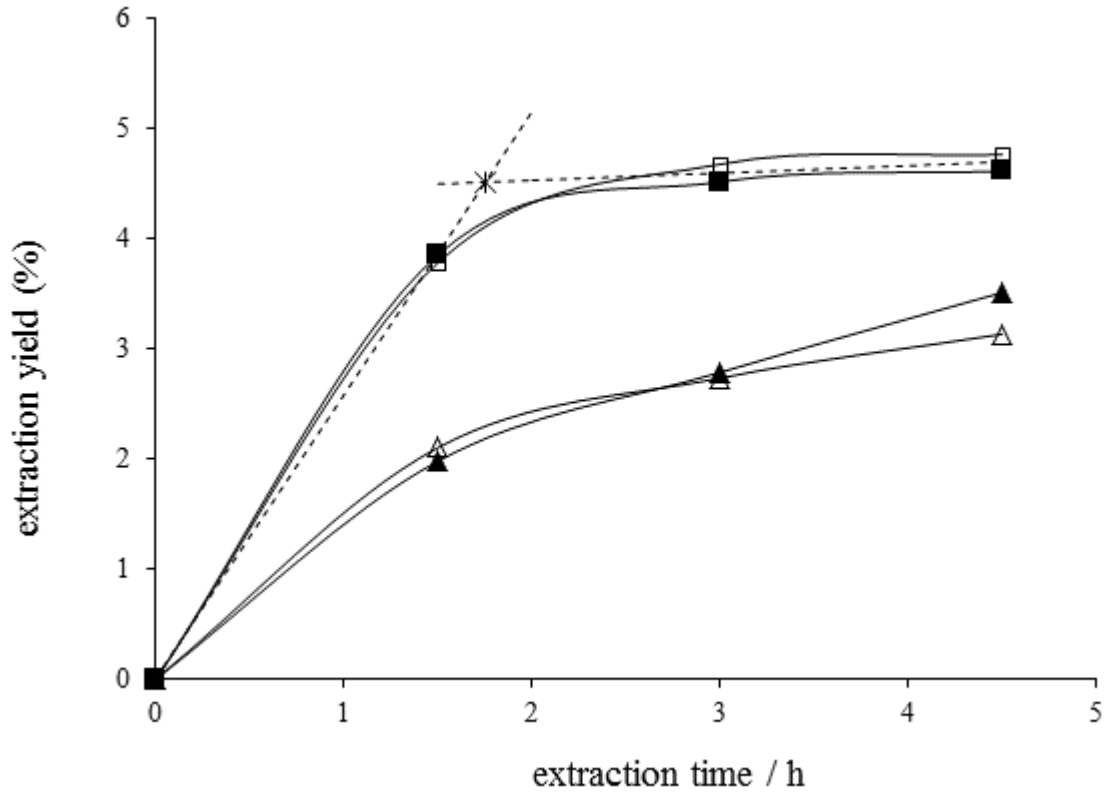
		t1	t2	t3
% weight thymol in oregano extracts				
	S1	0.55	0.28	-
	S2	10.36	7.97	1.92
% weight camphor in sage extracts				
	S1	4.65	1.36	0.61
	S2	17.28	2.08	0.91
% weight thymol in thyme extracts				
	S1	3.19	2.41	5.58
	S2	43.9	24.13	15.82
% weight carnosic acid in rosemary extracts				
	S1	12.03	15.54	19.05
	S2	1.82	7.55	12.30

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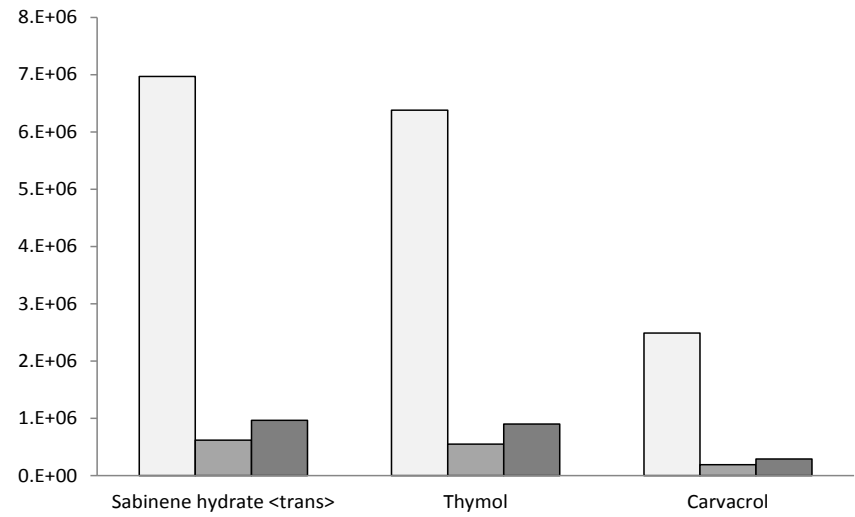
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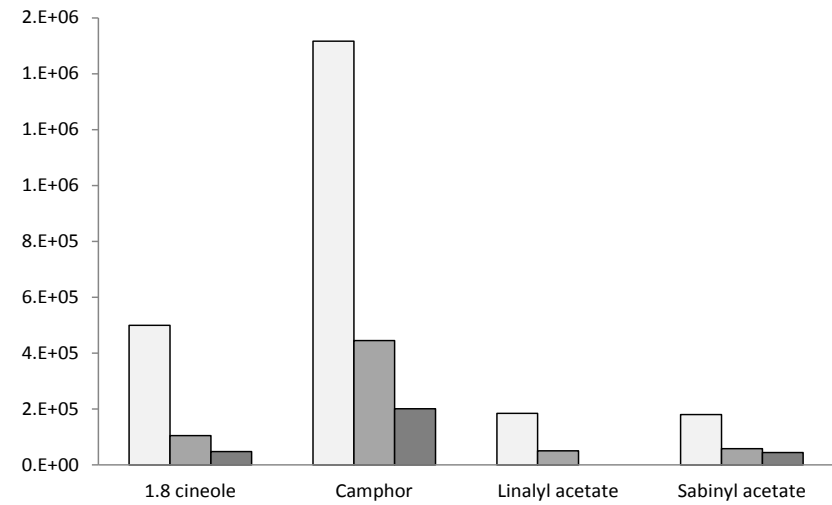


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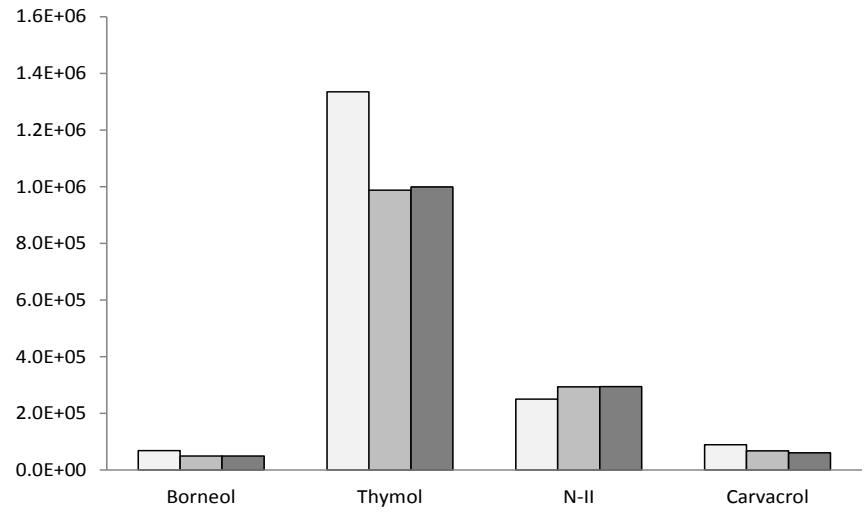
Figure 1. CO₂-SFE at constant pressure (30 MPa) of oregano (□), sage (■), thyme (△) and rosemary (▲). (*) Estimated optimal extraction time in the case of sage and oregano extraction.



(a) S1 - oregano extraction

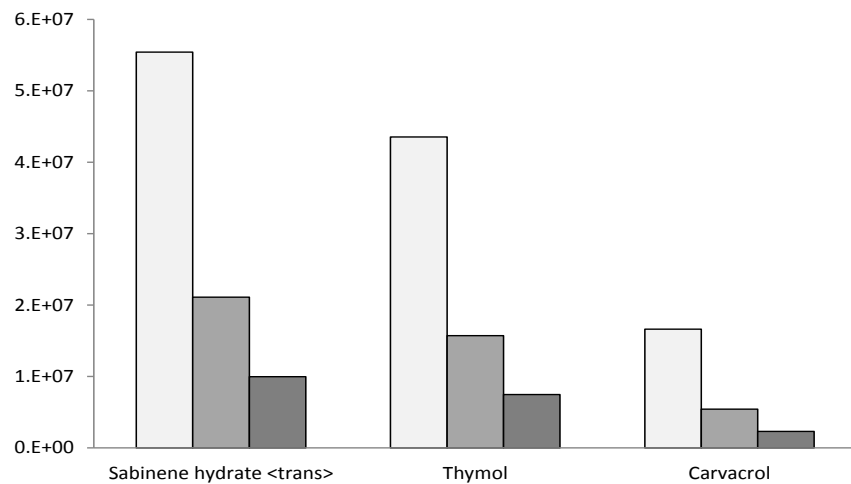


(b) S1 - sage extraction

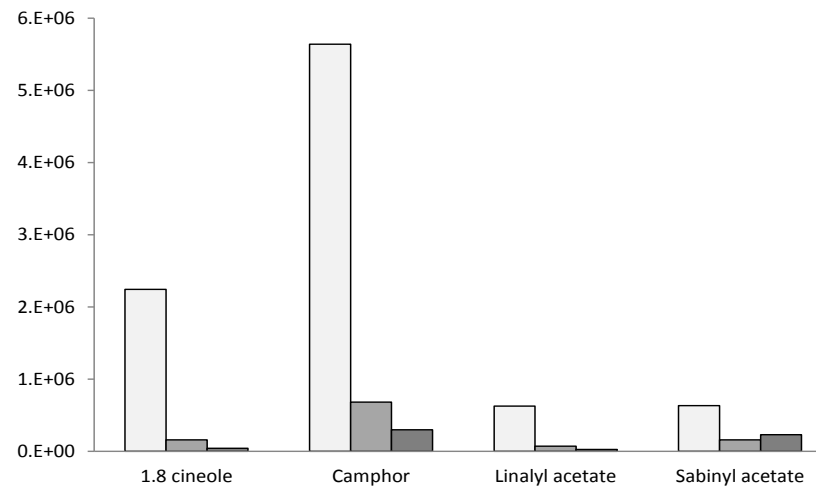


(c) S1 - thyme extraction

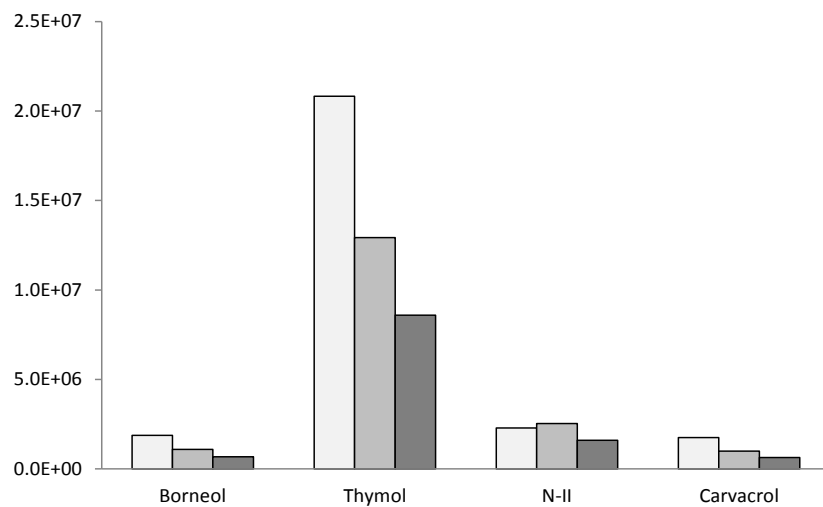
Figure 2. Kinetic behavior in the recovery of the main essential oil compounds identified in (a) oregano, (b) sage and (c) thyme S1 extracts.



(a) S2 – oregano extraction



(b) S2 – sage extraction



(c) S2– thyme extraction

Figure 3. Kinetic behavior in the recovery of the main essential oil compounds identified in (a) oregano, (b) sage and (c) thyme S2 extracts.

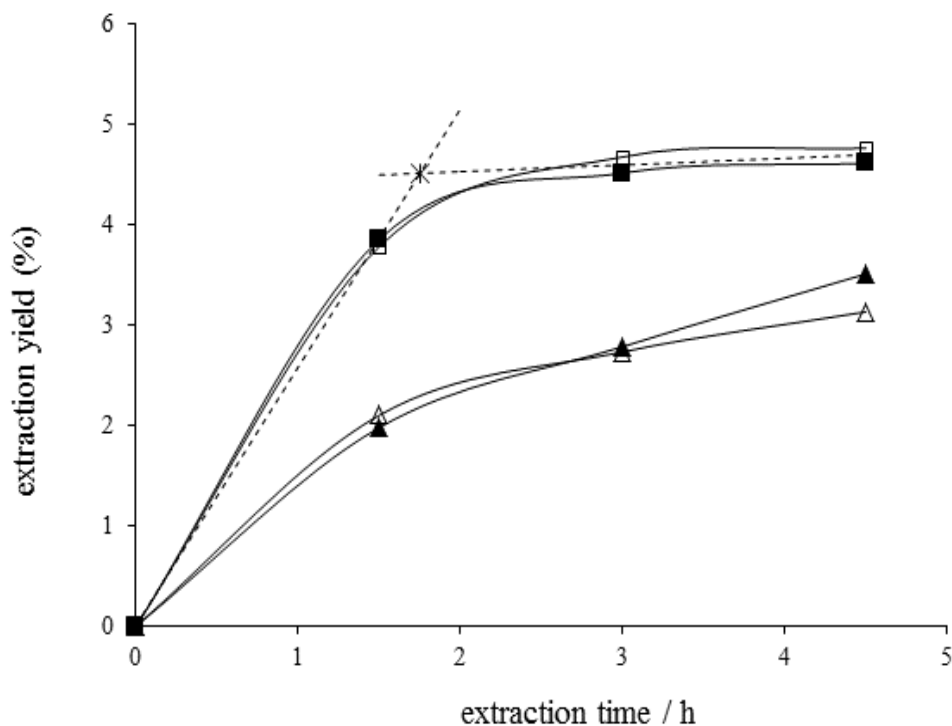
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Kinetic study of the supercritical CO₂ extraction of different plants from *Lamiaceae* family

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Supercritical CO₂ extraction of four different plants from *Lamiaceae* family, namely oregano (*Origanum vulgare*), thyme (*Thymus zygis*), sage (*Salvia officinalis*) and rosemary (*Rosmarinus officinalis*) was carried out in an experimental pilot-plant at 30 MPa and 313 K. Comparison of the kinetic performance reveals very similar behavior of oregano (□) and sage (■) extraction, so as for thyme (△) and rosemary (▲) extraction. A comparison between the extraction of the different plants was discussed, in terms of the extraction yields, the variation of the essential oil composition with time and the content of key bioactive substances identified in the different fractions.



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Highlights

- Supercritical CO₂ extraction of four different plants from *Lamiaceae* family was accomplished employing the same raw material pre-treatment, extraction and fractionation (two on-line decompressing separators) conditions.
- Comparison of the kinetic behavior reveals a removal velocity for thyme and rosemary almost half of the value corresponding to salvia and oregano.
- Oregano extract is mainly recovered in the first separator while the opposite behavior is observed for sage and thyme. In the case of rosemary extraction similar amounts of extract were recovered in both separators.
- Oregano, sage and thyme volatile oil compounds were selectively recovered (90.6, 77.6 and 93.4 %, respectively) in the second separator, while 72.4 % of the total antioxidant carnosic acid extracted from rosemary was selectively recovered in the first separator.
- The % weight of the monoterpene compounds (thymol and camphor) decrease with extraction time, while higher-molecular-weight compounds, such as phenolic diterpenes, increase with extraction time.