

Review

MAP3K kinases and kidney injury

Leticia Cuarental^{a,b,◇}, David Sucunza-Sáenz^{b,c,d,◇}, Lara Valiño-Rivas^{a,b},
Beatriz Fernandez-Fernandez^{a,b}, Ana Belen Sanz^{a,b}, Alberto Ortiz^{a,b},
Juan José Vaquero^{b,c,d,◇}, Maria Dolores Sanchez-Niño^{a,b,*,◇}

^a IIS-Fundacion Jimenez Diaz-UAM, Madrid, Spain

^b REDINREN, Spain

^c Departamento de Química Orgánica y Química Inorgánica, Universidad de Alcalá, 28871, Alcalá de Henares, Madrid, Spain

^d Instituto Ramón y Cajal de Investigación Sanitaria, (IRYCIS), Madrid, Spain

ARTICLE INFO

Keywords:

ASK1
MAP3K14
NIK
TWEAK
NF-kappaB
Kidney
MAP3K kinases
Diabetic kidney disease
Selonsertib
Acute kidney injury

ABSTRACT

Mitogen-activated protein kinases (MAP kinases) are functionally connected kinases that regulate key cellular process involved in kidney disease such as all survival, death, differentiation and proliferation. The typical MAP kinase module is composed by a cascade of three kinases: a MAP kinase kinase kinase (MAP3K) that phosphorylates and activates a MAP kinase kinase (MAP2K) which phosphorylates a MAP kinase (MAPK). While the role of MAPKs such as ERK, p38 and JNK has been well characterized in experimental kidney injury, much less is known about the apical kinases in the cascade, the MAP3Ks. There are 24 characterized MAP3K (MAP3K1 to MAP3K21 plus RAF1, BRAF and ARAF). We now review current knowledge on the involvement of MAP3K in non-malignant kidney disease and the therapeutic tools available. There is in vivo interventional evidence clearly supporting a role for MAP3K5 (ASK1) and MAP3K14 (NIK) in the pathogenesis of experimental kidney disease. Indeed, the ASK1 inhibitor Selonsertib has undergone clinical trials for diabetic kidney disease. Additionally, although MAP3K7 (MEKK7, TAK1) is required for kidney development, acutely targeting MAP3K7 protected from acute and chronic kidney injury; and targeting MAP3K8 (TPL2/Cot) protected from acute kidney injury. By contrast MAP3K15 (ASK3) may protect from hypertension and BRAF inhibitors in clinical use may induced acute kidney injury and nephrotic syndrome. Given their role as upstream regulators of intracellular signaling, MAP3K are potential therapeutic targets in kidney injury, as demonstrated for some of them. However, the role of most MAP3K in kidney disease remains unexplored.

© 2019 Sociedad Española de Nefrología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

* Corresponding author.

E-mail address: mdsanchez@fjd.es (M.D. Sanchez-Niño).

◇ These authors contributed equally.

<https://doi.org/10.1016/j.nefro.2019.03.004>

0211-6995/© 2019 Sociedad Española de Nefrología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

MAP3K quinasas y daño renal

R E S U M E N

Palabras clave:

ASK1
MAP3K14
NIK
TWEAK
NF-kappaB
Riñón
MAP3K quinasas
Enfermedad renal diabética
Selonsertib
Enfermedad renal crónica

Las proteínas quinasas activadas por mitógenos (MAP quinasas) son quinasas conectadas funcionalmente que regulan procesos celulares clave involucrados en la enfermedad renal como la supervivencia, la muerte, la diferenciación y la proliferación. El típico módulo MAP quinasa está compuesto por una cascada de 3 quinasas: una MAP quinasa quinasa quinasa (MAP3K) que fosforila y activa una MAP quinasa quinasa (MAP2K) que, a su vez, fosforila una MAP quinasa (MAPK). Si bien el papel de las MAPK como ERK, p38 y JNK se ha caracterizado bien en las lesiones renales experimentales, se sabe mucho menos acerca de las quinasas apicales en la cascada, las MAP3K. Hay 24 MAP3K (MAP3K1 a MAP3K21, más RAF1, BRAF y ARAF). En este trabajo revisamos el conocimiento actual sobre la participación de MAP3K en la enfermedad renal no maligna y las herramientas terapéuticas disponibles. Existe evidencia intervencionista experimental *in vivo* que respalda claramente el papel de MAP3K5 (ASK1) y MAP3K14 (NIK) en la patogenia de la enfermedad renal experimental. De hecho, el inhibidor de ASK1, selonsertib, ha sido estudiado en ensayos clínicos en la enfermedad renal diabética. Además, aunque la MAP3K7 (MEKK7, TAK1) es necesaria para el desarrollo renal, la inhibición de MAP3K7 en el adulto protegió de la lesión renal aguda y crónica experimental; e inhibir MAP3K8 (TPL2/Cot) protegió de la lesión renal aguda. Por el contrario, MAP3K15 (ASK3) puede proteger de la hipertensión y los inhibidores de BRAF, en uso clínico, pueden inducir lesión renal aguda y síndrome nefrótico. Dado su papel como reguladores de los primeros pasos de la señalización intracelular, las MAP3K son posibles dianas terapéuticas en la lesión renal, como se ha demostrado para algunas de ellas. Sin embargo, el papel de la mayoría de las MAP3K en la enfermedad renal no ha sido explorado.

© 2019 Sociedad Española de Nefrología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Chronic kidney disease (CKD) and acute kidney injury (AKI) are major causes of death and disability.^{1,2} It is estimated that 10% of the adult population worldwide has CKD while the incidence of AKI is growing.^{3,4} Both are associated with an increased risk of premature death and, in fact, CKD was among the fastest growing causes of death worldwide, according to the Global Burden of Disease (GBD) study.⁵ As an example, data from the GBD 2016 Spain analysis suggest that at the current rate of growth, CKD will become the second most common cause of death, after Alzheimer, by the year 2100.^{6,7} This growing burden of kidney disease reflects the need for earlier biomarkers of kidney injury that allow preventive rather than therapeutic intervention as well as the need for more effective therapeutic interventions.^{2,8} Kinases are a family of intracellular signaling regulators that have become therapeutic targets for diverse diseases and kinase inhibitors are in clinical use mainly in the oncology and immune regulation fields.⁹ Among kinases, mitogen-activated protein kinase kinase kinases (MAP3K) are of special interest given their localization at the apex of diverse intracellular signaling pathways in response to environmental stimuli and recent preclinical evidence supporting a key role of some of them, such as MAP3K14, in the pathogenesis of kidney injury.¹⁰ We now review MAP3Ks and the preclinical and clinical evidence suggesting a contribution to kidney injury and that they may become therapeutic targets in both AKI and CKD.

Mitogen-activated protein kinase cascades

Mitogen-activated protein kinases (MAP kinases) are functionally connected kinases that regulate key cellular process involved in kidney disease such as all survival, death, differentiation and proliferation. The typical MAP kinase module is composed by a cascade of three kinases: a MAP kinase kinase kinase (MAP3K) that phosphorylates and activates a MAP kinase kinase (MAP2K) which in turn phosphorylates a MAP kinase (MAPK) (Fig. 1). MAP3Ks are activated by MAP4Ks, oxidative stress, small proteins G or by TRAF adaptor proteins in response to growth factors, inflammatory cytokines, drugs, or physical stress. There are 24 characterized MAP3K, named from MAP3K1 to MAP3K21 plus RAF1, BRAF, and ARAF. Most of them are expressed by normal kidneys (Fig. 2).¹¹

The final targets of MAPK modules are transcription factors or other kinases that regulate gene expression by transcriptional or post-transcriptional mechanisms.¹² There are four well known and conventional MAPK pathways: the ERK pathway, the JNK pathway, the p38 pathway and the ERK5 pathway.^{13,14} The ERK pathway was the first characterized MAPK cascade. It is formed by three MAP3Ks RAF1, ARAF and BRAF, two MAP2Ks, MEK1 and MEK2 and two MAPK, ERK1 and ERK2, collectively known as ERK1/2. The ERK pathway has multiple functions, and is generally associated to survival and

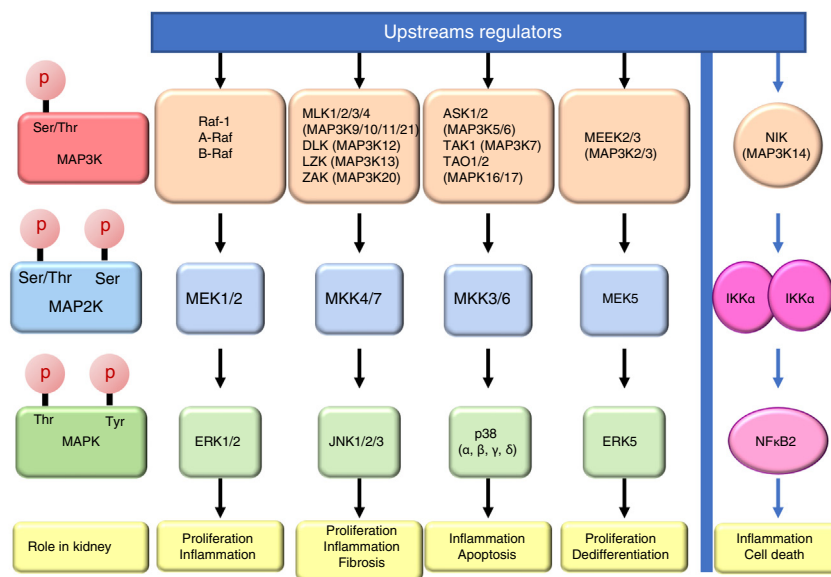


Figure 1 – The best characterized MAPK modules and cascades. Most information on a role of the cascade in the kidney was obtained by studying the MAPK component of the cascade and it cannot be extrapolated to the MAP3K component since these enzymes may have additional functions.

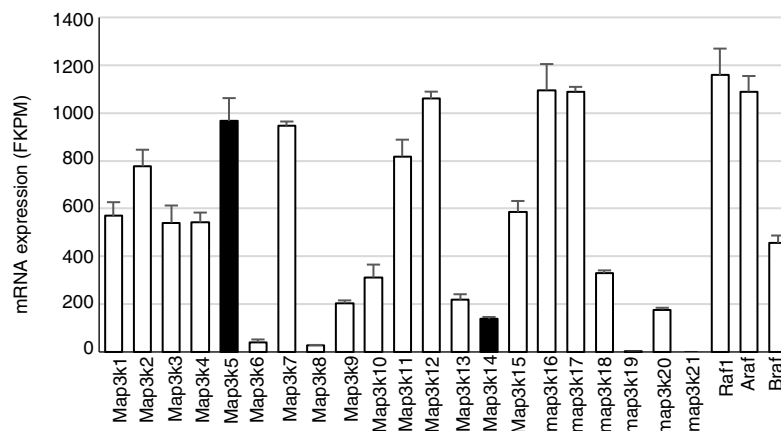


Figure 2 – Constitutive MAP3K expression in murine kidneys. Data obtained by RNA-seq analysis of three normal murine kidneys. Mean \pm SD of gene expression in kidneys from 3 wild-type control mice.¹¹

proliferation in different tissues including kidney.^{15,16} However, additional functions have been described. As examples, in the kidney ERK pathway mediated TWEAK-induced proliferation and pro-inflammatory response in tubular cells.^{15,17} The JNK pathway is composed by seven MAP3K, MLK1–4 (respectively MAP3K9–11 and MAP3K21), DLK (MAP3K12), LZK (MAP3K13) and ZAK (MAP3K20); two MAP2Ks (MKK4/7) and three MAPKs (JNK1, JNK2, JNK3). It is generally related to cell proliferation, and it has also been linked to the development of renal fibrosis and inflammation in cyclosporine nephrotoxicity.^{18–21} The p38 pathway is formed by 5 MAP3Ks, ASK1/2 (MAP3K5/6 respectively), TAK1 (MAP3K7), TAO1 (MAP3K16), and TAO2 (MAP3K17); two MAP2Ks MKK3/6 and four MAPKs p38 (α , β , γ , δ). This pathway regulates and

frequently promotes renal inflammation and apoptosis.^{22,23} As examples, p38 mediates proliferation of tubular cells induced by TWEAK and expression of inflammatory mediators in renal cells induced by MIF.^{15,24} The ERK5 pathway is composed by 2 MAP3K, MEEK2/3 (MAP3K2 and 3 respectively); MAP2K MEK5 and MAPK ERK5. ERK5 is the only ERK protein that contains a nuclear localization signal it appears to regulate cell proliferation and dedifferentiation in the kidney, like ERK1/2.^{18,25,26}

ERKs 3, 4, 7 are atypical pathways and the upstream MAPKs involved in their activation and function are poorly characterized, and some evidences indicate that could be activated by autophosphorylation.¹⁸ Moreover, ERK 3 and 4 could also be activated by Group I p21-activated Kinases (PAKs) indicating

that they are not activated by a typical MAPK module.²⁷ ERK8 is the most recently identified protein kinase that is regulated by autophosphorylation and specific upstream activating kinase are unknown.²⁸

Additional MAP3K are not associated with specific MAPK cascades. One example is NF- κ B-inducing kinase (NIK), initially identified as a kinase that participates in NF κ B activation. The amino-acid sequence revealed that it is a serine/threonine kinase like MAP3Ks, although the signaling pathway is not a classical MAPKs module.²⁹ NIK is a key protein for non-canonical, NF κ B2 activation and is required for the physiological immune response and also promotes inflammation. In the kidney, NIK targeting may be protective during kidney injury.^{10,30}

Drugs targeting MAP3K kinases

A search in Scifinder (<https://scifinder.cas.org>) for small molecule inhibitors of MAP3K, made by searching individual molecules by their official gene symbol and alternative names, disclosed that small molecule inhibitors were under study for 14 of them (Table 1). Inhibitors for five MAP3K are in clinical use for malignancy, although in some cases MAP3K inhibition is a promiscuous effect of a drug targeted to a different kinase.

MAP3K

Table 2 summarizes current knowledge about MAP3K involvement in non-malignant kidney disease. Our group has been instrumental in developing the evidence base supporting the involvement of MAP3K14/NIK as a pathogenic molecule in kidney disease. However, the potential role of most MAP3K in kidney disease remains unexplored in vivo.

MAP3K14/NIK

MAP3K14/NIK has been conclusively demonstrated to be activated in experimental kidney disease and to contribute to the severity of experimental kidney injury (Fig. 3). NIK is the essential upstream serine/threonine kinase of the non-canonical NF κ B pathway.²⁹ It is now known that NIK protein levels are regulated both transcriptionally (as in AKI and in tubular cells in response to TWEAK) and post-transcriptionally.¹⁰ Thus, concentrations are low in quiescent cells because of low mRNA levels and rapid degradation of the protein. Cytokines and oxidative stress may increase NIK mRNA as well as protein stability, leading to NIK activation.^{10,31} NIK induces I κ B kinase- α (IKK- α)-mediated phosphorylation of NF κ B2 p100, and phosphorylated p100 undergoes ubiquitination and subsequent proteasomal processing to active NF κ B2 p52.³² Efficient NF κ B2 p100 ubiquitination requires Uba3 (ube1c) and Ube2m (UBC12), ube2d3 (UBCH5c) and intact Cullin1 in SCF β -TrCP.³³ Interestingly, the Ube2m E2 ubiquitin conjugating enzyme and cullin-1 of the SCF β -TrCP E3 ubiquitin ligase were upregulated in AKI.¹⁰ Additional functions of NIK have been recently reported, but their role in kidney injury remains unclear.^{34–37}

NF κ B promotes inflammation in the kidney, as well as having multiple other functions such as promoting cell

survival or decreasing the expression of anti-inflammatory proteins such as Klotho and PGC1 α in response to TWEAK or tissue injury, while the involvement of NF κ B in other TWEAK-dependent responses such as induction of kidney-protective CCL20 expression has not been characterized.^{38–44} Recent studies have focused on NF κ B activity modulation by proteins such as Bcl3 and NF κ Biz.^{45,46} Canonical NF κ B activation, a rapid process triggered by multiple stimuli has been best characterized and is responsible for most activities of TWEAK in cultured tubular cells, including expression of most chemokines⁴⁷ although some (e.g. CCL21) were early on show to be dependent on non-canonical NF κ B activation.³⁰ Although non-canonical NF κ B activation has been reported in several kidney disease, the pathogenic role of NIK has been best characterized in experimental AKI induced by folic acid (crystal nephropathy) or cisplatin.^{30,48,49} Evidence for NIK activation in vivo in AKI included increased NIK mRNA and protein levels, NF κ B p100 processing to NF κ B p52, increased nuclear localization and DNA binding activity of p52/RelB and decreased kidney inflammation, cell death and fibrosis and preserved renal function in NIK activity deficient mice, which were protected from AKI (Fig. 2).¹⁰ Upregulation of tubular cells NIK was observed in both human AKI and CKD, suggesting that NIK maybe a therapeutic target in the clinical settings. Protection from AKI may depend on systemic NIK deficiency (e.g. leukocyte NIK deficiency) and/or kidney NIK deficiency. Cell culture and bone marrow transplantation experiments support the hypothesis that renal cell NIK targeting is necessary for nephroprotection. NIK and non-canonical NF κ B gene targets have been characterized in the immune system, but there was little prior information on kidney cells.³⁸ In this regard, NIK siRNA targeting reduced inflammatory responses and serum deprivation-induced death in cultured tubular cells.¹⁰ Interestingly, NIK targets in tubular cells included both known non-canonical (e.g. CCL21, CXCL10) and canonical (MCP1, Rantes) NF κ B pathway targets.^{10,30} These results were consistent with decreased inflammation and tubular cell apoptosis in vivo in NIK activity-deficient mice during AKI. These results are also consistent with observations targeting another component of the non-canonical NF κ B pathway, RelB. Thus, RelB targeting by siRNA protected mice against lethal kidney ischemia-reperfusion, and RelB targeting in cultured proximal tubular cells, protected from apoptosis induced by TNF-cisplatin.^{50,51}

Since NIK deficiency results in immune deficiency a potential role of immune cell NIK in kidney injury cannot be completely excluded. Thus, NIK inhibitors will be expected to target both renal cells and leukocytes and may have adverse effects related to immune suppression. The successful use of NIK inhibitors in experimental systemic lupus erythematosus, and, specifically, for lupus nephritis, should be viewed in the context of their immunosuppressive properties.⁵² Thus, systemic lupus erythematosus is an autoimmune kidney disease and the current standard of therapy for lupus nephritis is immune suppression.

Non-canonical NF κ B activation also contributes to podocyte injury induced by TWEAK activation of this receptor Fn14.^{49,53} In cultured podocytes, TWEAK increased the expression of the chemokines CCL21, CCL19 and RANTES. The CCL19 and the early RANTES responses depended on canon-

Table 1 – MAP3K inhibitors. List according to a search in Scifinder (<https://scifinder.cas.org>) by their official gene symbol and alternative names performed on September 15, 2018. The same table with references can be found as 1.

Gene symbol (name)	Inhibitors	Representative inhibitor	Inhibitors in clinical use
MAP3K1 (MEKK1)	No		No
MAP3K2	Yes		No
MAP3K3 (MEKK3)	No		No
MAP3K4	No		No
MAP3K5 (ASK1)	Yes	GS-4997 (Phase 2)	No
MAP3K6 (ASK2)	No		No
MAP3K7 (MEKK7, TAK1)	Yes	5Z-7-oxozeaenol	No
MAP3K8 (TPL2)	Yes		No
MAP3K9	No		No
MAP3K10	No		No
MAP3K11 (MEKK11, MLK3)	Yes	URMC-099 – –	No
MAP3K12 (MUK, DLK)	Yes	– CEP-1347 GNE-3511 GNE-8505	Sunitinib (Sutent, SU11248)
MAP3K13 (LZK)	Yes	–	No
MAP3K14 (NIK)	Yes	– NIK SMI1 Patent	No
MAP3K15 (ASK3)	No	Patent –	No
MAP3K16 (TAO1, TAOK1)	Yes	Patent	No
MAP3K17 (TAO2, TAOK2)	Yes	Patent –	No
RAF1 (CRAF)	Yes		Regorafenib (Stivarga) Inhibition of RAF-1 > BRAF Sorafenib (Nexavar; Sorafenibum; BAY 43-9006) Inhibition of RAF1 > BRAF Vemurafenib (Zelboraf, PLX4032/RG7204/RO5185426), inhibition of RAF1 ~BRAF Dabrafenib (Tafinlar, GSK2118436) inhibition BRAF>RAF1 Lifirafenib (BGB-283) Inhibition RAF1 > BRAF GW5074 PLX-4720 AZ628 ZM 336372 ZM 336372 GW 5074 RAF265 NVP-BHG712 TAK-632 RAF709 CCT196969 CCT241161 BAW2881 (NVP-BAW2881) LY3009120
BRAF	Yes	SB-590885/SB-699393 AZ304 GDC-0879 CEP-32496 RO5126766 (CH5126766) LY3009120	Encorafenib (Braftovi, LGX818) selectivity for BRAF
ARAF	Yes		Lifirafenib (BGB-283) Inhibition A-RAF > RAF1 > BRAF

– Table 1 (Continued)

Gene symbol (name)	Inhibitors	Representative inhibitor	Inhibitors in clinical use
ZAK (MLT, MLTK, HCCS-4, MRK, AZK)	Yes	PLX-4720 AST-487	Sorafenib Vemurafenib Imatinib (Gleevec) Nilotinib (Tasigna) Ponatinib (Iclusig) Motesanib/AMG-706 Bofutinib/SKI-606 (Bosulif)

Table 2 – MAP3K and non-malignant kidney injury.

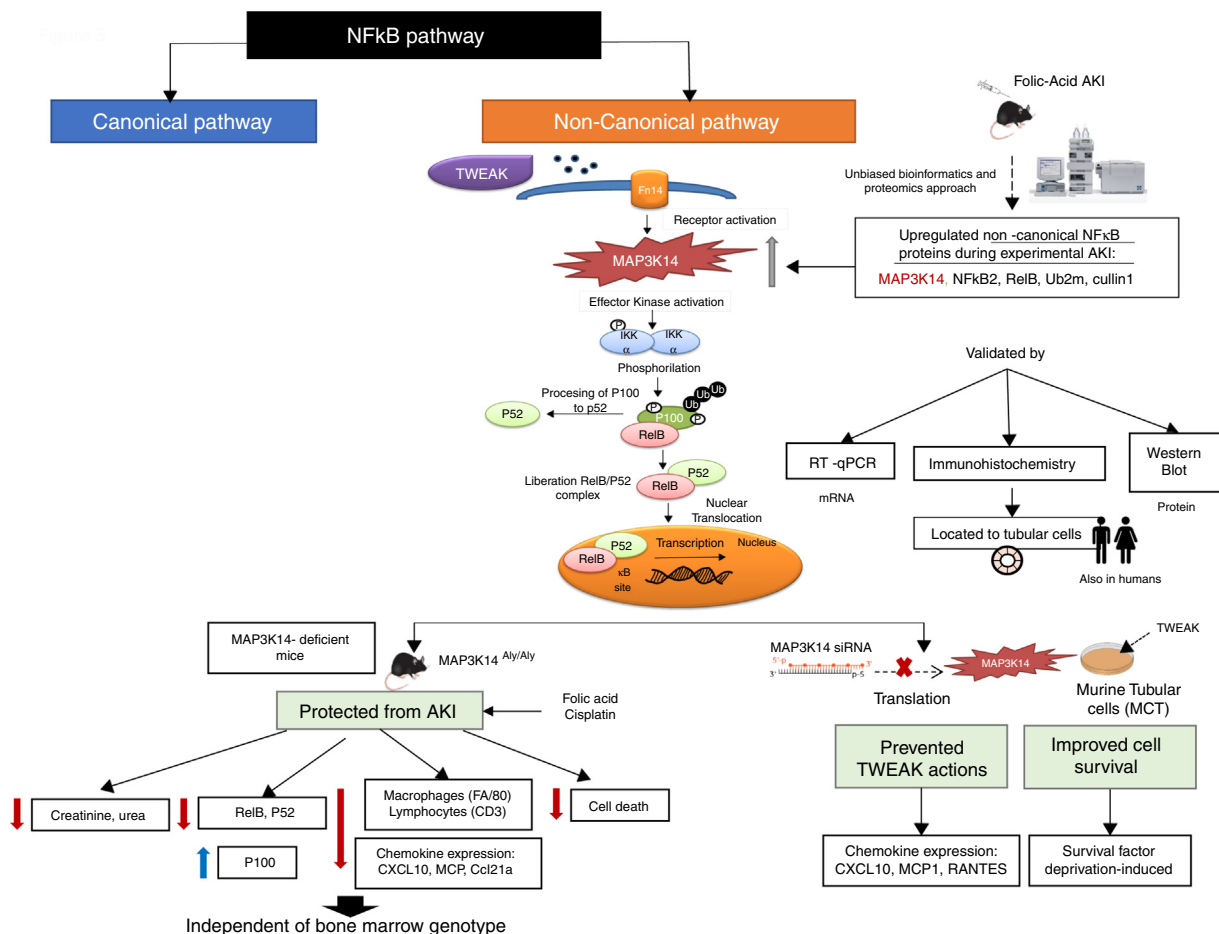
Gene symbol (name)	Expression/activation in kidney injury	Targeting in kidney injury	Ref.
MAP3K1 (Mekk1)		Protective? Mekk1 directly represses PKD1 transcription in cultured cells. No in vivo studies	[112]
MAP3K5 (ASK1)	Increased: activity in glomerular and/or tubular epithelial cells in IRI-AKI, DKD, glomerulopathies (crescentic glomerulonephritis/experimental membranous nephropathy) and kidney fibrosis induced by UUO	Protective. Targeting Ask1 with inhibitors and/or genetic deficiency protective in AKI, glomerular injury (crescentic glomerulonephritis), diabetic kidney disease and kidney fibrosis (UUO). Clinical trials in DKD did not meet endpoints.	[63–65,68,70,71,113]
MAP3K7 (MEKK7, TAK1)	Increased: TAK1 increased in cisplatin-induced AKI, but not in UUO in mice.	Protective: Conditional TAK1 ^{−/−} protected from kidney fibrosis (UUO); TAK1 inhibition protected from cisplatin-induced or IRI AKI and IRI fibrosis.	[77–79]
MAP3K8 (TPL2)	Cot/Tpl2 phosphorylated in IRI	Deleterious: MAP3K7 ^{−/−} spontaneous neonatal kidney scarring with retention of embryonic nephrogenic rests. Protective: Cot/Tpl2 ^{−/−} mice protected from IRI: decreased apoptosis	[80] [81]
MAP3K14	MAP3K14 upregulation in folate-induced AKI	Protective: MAP3K14 KO protected from folate and cisplatin-induced AKI	[10]
MAP3K15 (ASK3)		Deleterious: hypertensive phenotype of ASK3 ^{−/−} mice	[82]
BRAF (v-RAF)		Deleterious: BRAF inhibitor nephrotoxicity in humans: AKI and podocyte injury	[104,106]

AKI: acute kidney injury; IRI: ischemia/reperfusion injury; DKD: diabetic kidney disease; UUO: unilateral ureteral obstruction

ical NFκB activation, while the CCL21 response depended on non-canonical NFκB activation and could be inhibited by a specific NIK siRNA. Increased kidney Fn14 and CCL21 expression was also observed in podocytes in rat proteinuric kidney disease induced by puromycin, supporting the in vivo relevance of the findings.⁴⁹ In this regard, there is experimental and human kidney disease evidence supporting a role of TWEAK/Fn14 in promoting podocyte injury in proteinuric kidney disease.⁵³ The potential contribution of NIK to fibroblast proliferation and activation induced by TWEAK and to kidney fibrosis^{54,55} or in endothelial cell responses to TWEAK⁵⁶ has not been explored. In this regard, TWEAK, as other inflammatory cytokines, promotes vascular calcification and there is evidence of the cooperation between canonical and

non-canonical NFκB signaling, since a siRNA targeting RelB reduced by 20% TWEAK pro-calcific effects and decreased TWEAK-induced loss of human vascular smooth muscle cells contractile phenotype and MMP9 activity, while other effects were prevented by agents blocking canonical activation.^{57,58}

The p52/RelB heterodimers characteristic of NIK-initiated non-canonical NFκB activation share a number of gene targets with RelA-containing, classically activated NFκB complexes.^{59–61} Since canonical NFκB activation is an early transient response peaking at around 1–3 h while and non-canonical NFκB activation is delayed and peaks at around 24 h, non-canonical NFκB activation may contribute to sustained NFκB-dependent gene expression.^{38,62}



ASK1/MAP3K5

Apoptosis signal-regulating kinase 1 (ASK1) is activated by oxidative stress and, in turn, activates MAPK p38 and JNK and induces apoptotic, inflammatory, and fibrotic signaling in settings of oxidative stress.²³ There is convincing evidence of the pathogenic role of ASK1 in experimental kidney disease derived from the study of specific ASK1 inhibitors or ASK1 deficient mice and a clinical trial focused on diabetic kidney disease (DKD) (Table 2).

The first reports focused on AKI. Ask1^{-/-} mice are healthy and are protected from AKI induced by ischemia-reperfusion injury (IRI), while Ask1^{-/-} cultured tubular cells are protected from hypoxia-induced cell death and transfection with dominant-active ASK1 induced apoptosis.^{63,64} Additionally, transfection of tubular cells with dominant-active ASK1 induced apoptosis decreased chemokine production. Evidence from Ask1 involvement in AKI is also derived from studies with small molecule inhibitors. NQDI-1, an ASK1 inhibitor attenuated renal dysfunction and histological changes in rats with IRI-AKI.⁶⁵ In the same model, Oral GS-444217 reduced serum creatinine and urea, tubular cell death and proinflammatory and profibrotic mediators in the kidney.²³ GS-444217 is

a selective ATP-competitive small-molecule inhibitor of ASK1 that inhibits ASK1 autophosphorylation and ASK1-mediated signaling in cells and in vivo.²³ Nephroprotection may expand to additional kidney diseases characterized by tubular injury, such as myeloma kidney, which is triggered by the presence in urine of massive amounts of immunoglobulin free light chains. Thus, in cultured human proximal tubular cells, free immunoglobulin light chains increased Ask1 phosphorylation and Ask1 siRNA targeting protected tubular cells from light chain-induced apoptosis.⁶⁶

Protection by Ask1 targeting has also been reported for experimental glomerular injury. In glomerular epithelial cells (GEC), sublytic complement activation induced oxidative stress and activated Ask1, JNK and p38 MAPK and this was decreased by a dominant negative ASK1 mutant. ASK1 promoted complement-mediated lysis and this was dependent on p38 MAPK activity.⁶⁷ The in vivo relevance of these findings was supported by the observation of increased glomerular ASK1 activity during experimental membranous nephropathy, a proteinuric kidney disease characterized by podocyte (glomerular epithelial cell).⁶⁷ However, in vivo targeting was not attempted. More recent reports explored Ask1 inhibitors. In accordance to prior observations of MAPK p38 and JNK activation and the beneficial effect of their targeting

in experimental crescentic glomerulonephritis, GS-444217 decreased glomerular inflammation and fibrosis, proteinuria and preserved renal function in diverse rat experimental models of crescentic glomerulonephritis.⁶⁸

Unilateral ureteral obstruction (UUO) is used as a model of accelerated CKD that recapitulates most of the human features of the disease.⁶⁹ p38 and JNK were activated in kidney fibrosis induced by UUO. In Ask1^{-/-} mice with ureteral obstruction, p38 was not activated, JNK activation was decreased and kidney inflammation and fibrosis were milder. The key cell targets responsible for kidney protection appeared to be the tubular cells, since tubular cell p38 activation was observed in vivo and Ask1-deficient cultured tubular cells were protected from angiotensin II and H₂O₂, induced p38 activation and upregulation of TGFβ1, and chemokines, while Ask1^{-/-} fibroblast activity was not impaired.⁷⁰

However, the studies with more translational potential explored DKD. The ASK1 pathway is activated in glomeruli and tubules of human and experimental DKD. GS-444217 reduced inflammation and fibrosis, prevented albuminuria development and preserved renal function in experimental DKD and the effect was observed on top of renin-angiotensin system blockade.²³ However, in earlier studies in streptozotocin-induced diabetes in hypertensive endothelial nitric oxide synthase (Nos3)-deficient mice, GS-444217 did not decrease hypertension or albuminuria, but decreased diabetic glomerulosclerosis and inflammation and reduced renal function.⁷¹ A Phase 2 placebo-controlled clinical trial investigating the effects of three different doses of selonsertib (GS-4997) on eGFR (primary end-point) and albuminuria (secondary end-point) at week 48 in patients with type 2 diabetes was completed in 2016.⁷² The fact that in a phase 2 RCT the primary endpoint was ambitious, impact on eGFR and not albuminuria is unusual⁷³ and likely reflects the lack of impact of Ask1 inhibition in at least some experimental models of DKD.⁷¹ While the results of the clinical trials have not been reported in full, Gilead Sciences announced in 2016 that selonsertib did not meet endpoints in DKD or pulmonary arterial hypertension, but clinical development continues with active phase 3 trials for nonalcoholic Steatohepatitis (NASH: NCT03053063, NCT03053050) that completed enrollment in 2018.⁷⁴ Although as of December 6, 2018 there is no planned or ongoing selonsertib trial in DKD in clinicaltrials.gov, clinical development for kidney disease appears to continue since phase 1 RCT were registered in the Australia New Zealand Clinical Trial Registry (ANZCTR) and other countries for patients with CKD in the summer of 2018.⁷⁵ Furthermore, at the 2018 American Society of Nephrology Renal Week, a post hoc analysis of the phase 2 DKD trial was presented.⁷⁶ The analysis excluded 2 sites for technical deviations as well as patients with baseline eGFR <20 mL/min/1.73 m². These exclusions may represent the identification of issues that blurred the information provided by the trial and considered not promising in 2016. In this post hoc analysis, the highest dose of selonsertib (18 mg) was associated with significantly milder eGFR slopes than placebo both in the overall trial population and in a high risk population defined by high baseline soluble TNFR1 (sTNFR1) levels.⁷⁶

Other MAP3K kinases: MAP3K7, MAP3K8 and MAP3K15

Additional MAP3K have been targeted in vivo in experimental kidney disease (Table 2). MAP3K7 (MEKK7, TAK1) was increased in cisplatin-induced AKI, but not in UUO in mice.^{77–79} However, for kinases activity is more relevant than levels of expression. In this regard, conditional TAK1^{-/-} mice were protected from kidney fibrosis (UUO) and TAK1 inhibition with small molecules protected from cisplatin-induced or IRI AKI and IRI fibrosis.^{77–79} Specifically, in UUO, TAK1 deletion resulted in suppression of JNK, p38, and NF-κB activation, and milder inflammation and fibrosis.⁷⁷ Despite this success, caution should be exercised in the extrapolation of the results to the clinic, since MAP3K7 appears to have a key role in kidney development and developmental genes may be re-expressed during kidney disease and contribute to kidney repair. Thus, MAP3K7^{-/-} mice develop spontaneous neonatal kidney scarring with retention of embryonic nephrogenic rests.⁸⁰

There is also some experience targeting other MAP3K in vivo. MAP3K8 (TPL2/Cot) is phosphorylated during IRI and Cot/Tpl2^{-/-} mice were protected from IRI and apoptosis was decreased.⁸¹ By contrast MAP3K15 (ASK3) may have a role in protecting from hypertension. Thus, ASK3^{-/-} mice have a hypertensive phenotype.⁸²

Lessons from kinase inhibitors in clinical use

Current available treatments for CKD are based on renal angiotensin-system (RAS) blockade and immunosuppressive drugs and can delay the development of renal failure but inflammation and fibrosis keep progressing.^{73,83,84} Several orally available kinase inhibitors, are currently in the pipeline for the kidney disease.⁶³ Data obtained from the clinical use of kinase inhibitors outside nephrology may help to define the role of target kinases in the human context, be it either because of the development of adverse effects or, as described for the Nrf2 activator bardoxolone, because kidney function or adverse events are observed to improve.^{83,85} In any case, nephrotoxicity observed with the high doses used to treat cancer does not preclude a potential therapeutic effect of a lower dose in kidney disease, although it does provide a note of caution.

Since the approval of Imatinib, in 2001, more than 30 small-molecule kinase inhibitors are being used or tested in cancer treatment^{86–89} and inflammatory diseases.^{90,91} Most of the approved kinase inhibitors are used to treat malignancy.^{87,92–94} or, as is the case for JAK inhibitors baricitinib and tofacitinib, autoimmune disease.^{95,96} Baricitinib also reduced albuminuria in patients with DKD.^{73,97} Some kinase inhibitors are nephrotoxic. The most consistent nephrotoxicity is associated with drugs targeting the VEGF pathway and its tyrosine kinase receptor VEGFR, which can induce hypertension, albuminuria, nephrotic syndrome or even thrombotic microangiopathy, and this is consistent with preclinical advances on its role in endothelial and podocyte well-being.^{98,99} Some kinase inhibitors are promiscuous and, thus it is difficult to related

side effects with inhibition of a specific kinase. This is the case for sorafenib, which inhibits VEGFR, PDGFR and BRAF, among others.

There are fewer drugs in clinical use targeting MAP3K. The ones available may inhibit several MAP3K, and thus, it is difficult to pinpoint the exact kinase responsible for any kidney actions. However, nephrotoxicity has been reported for drugs that among other targets, inhibit BRAF (Table 1). Vemurafenib and dabrafenib are BRAF inhibitors approved for treatment of late-stage melanoma.¹⁰⁰ Vemurafenib has been associated to the development of AKI. Decreased of renal function (mean reduction of 29 ml/min) was associated with nephrotic range proteinuria in 5 patients.^{101,102} A subsequent study in 74 patients treated with vemurafenib, found severe but reversible AKI predominantly in men with proven tubular and interstitial damage in renal biopsy within the first 3 months of treatment.¹⁰³ In the FDA review on Adverse Event Reporting Systems of Vemurafenib, there were 132 cases of AKI, 85 in men, and 47 in women.¹⁰⁴ A recent report observed AKI in 25% of melanoma patients treated with a combination therapy of the MEK inhibitor cobimetinib and vemurafenib, which was 60% lower than in prior results with vemurafenib monotherapy.¹⁰⁵ Dabrafenib also has nephrotoxic potential, with 13 cases of AKI detected from April 2013 to June 2014, mostly in men (12 men vs 1 woman).¹⁰⁴ Podocyte injury leading to nephrotic developed in one patient during dabrafenib and trametinib treatment.¹⁰⁶ In cultured podocytes, BRAF inhibition decreased PLC ϵ 1 and nephrin expression, inhibited the podocyte VEGF system and increased the permeability to albumin.¹⁰⁶ A patient on encorafenib and binimetinib, BRAF and MEK inhibitor, respectively, developed after two months renal dysfunction and proteinuria due to crescentic glomerulonephritis with a granulomatous reaction which reversed following withdrawal of the chemotherapy.¹⁰⁷ In any case, although clinical nephrotoxicity is observed when used for malignancy, it is still worth testing these agents for experimental kidney disease in case a lower dose may favorably impact kidney injury. We should remember that the first reports on captopril and the kidney referred to AKI and glomerular injury at a time when considerably higher doses were used.^{108–111}

Conclusions

MAP3K sit at the cusp of intracellular signaling cascades, making them potential targets for therapeutic intervention. However, for the very same reason they may impact multiple cell processes. They may be targeted by small molecules, but the similarity between them is hindrance to the development of specific inhibitors. Most of them remain unexplored from the kidney point of view and this is an opportunity. Human experience is for the most part limited to small molecules used in cancer therapy. In this regard, glomerular and tubular nephrotoxicity has been described, especially for BRAF inhibitors. However, there is experimental evidence derived from genetically modified mice and/or the in vivo use of small molecule inhibitors for a pathogenic role of MAP3K14/NIK, ASK1/MAP3K5, MAP3K7/MEKK7/TAK1, and MAP3K8/TPL2/Cot in AKI and/or CKD. Even a clinical trial of

ASK1/MAP3K5 targeting was performed in DKD, although the results have not been published. Given the safety up to now of ASK1/MAP3K5 inhibitors in humans and the wide range of responsive experimental nephropathies to MAP3K14/NIK, ASK1/MAP3K5, MAP3K7/MEKK7/TAK1, and MAP3K8/TPL2/Cot, further research should be aimed at defining the optimal drug, dose and disease target for human drug development as well as to define if there is any pathogenic role in kidney disease for those MAP3K still unexplored in the kidney.

Conflict of interest

The authors declare no conflict of interest.

Acknowledgments

Grant support: Instituto de Salud Carlos III (ISCIII) and FEDER – Fonds Européen de Développement Économique et Régional (FEDER) funds EUTOX, CP14/00133, PI15/00298, PI16/01900, PI16/02057, PI18/01366, Sociedad Española de Nefrología, Fundación Renal Íñigo Álvarez de Toledo (FRIAT), ISCIII Red de Investigación Renal (REDinREN) RD016/009, Salary support: ISCIII Miguel Servet to MDSN, Universidad Autónoma de Madrid to LC.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.nefro.2019.03.004](https://doi.org/10.1016/j.nefro.2019.03.004).

REFERENCES

- Ortiz A, Covic A, Fliser D, Fouque D, Goldsmith D, Kanbay M, et al. Epidemiology, contributors to, and clinical trials of mortality risk in chronic kidney failure. *Lancet* (London, England). 2014;383:1831–43, [http://dx.doi.org/10.1016/S0140-6736\(14\)60384-6](http://dx.doi.org/10.1016/S0140-6736(14)60384-6).
- Sanchez-Niño MD, Sanz AB, Ramos AM, Ruiz-Ortega M, Ortiz A. Translational science in chronic kidney disease. *Clin Sci* (Lond). 2017;131:1617–29, <http://dx.doi.org/10.1042/CS20160395>.
- Thomas B, Matsushita K, Abate KH, Al-Aly Z, Ärnlöv J, Asayama K, et al. Global cardiovascular and renal outcomes of reduced GFR. *J Am Soc Nephrol*. 2017;28:2167–79, <http://dx.doi.org/10.1681/ASN.2016050562>.
- Otero A, de Francisco A, Gayoso P, García F. EPIRCE Study Group Prevalence of chronic renal disease in Spain: results of the EPIRCE study. *Nefrología*. 2010;30:78–86, <http://dx.doi.org/10.3265/Nefrología.pre2009.Dic.5732>.
- GBD 2016 Causes of Death Collaborators. Global, regional, and national age-sex specific mortality for 264 causes of death, 1980–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet* (London, England). 2017;390:1151–210, [http://dx.doi.org/10.1016/S0140-6736\(17\)32152-9](http://dx.doi.org/10.1016/S0140-6736(17)32152-9).
- Soriano JB, Rojas-Rueda D, Alonso J, Antó JM, Cardona P-J, Fernández E, et al. The burden of disease in Spain: results from the Global Burden of Disease 2016. *Med Clin* (Barc). 2018;151:171–90, <http://dx.doi.org/10.1016/j.medcli.2018.05.011>.

7. Ortiz A, Sanchez-Niño MD, Crespo-Barrio M, De-Sequera-Ortiz P, Fernández-Giráldez E, García-Maset R, et al. The Spanish Society of Nephrology (SENEFRO) commentary to the Spain GBD 2016 report: keeping chronic kidney disease out of sight of health authorities will only magnify the problem. *Nefrologia*. 2019;39:29–34, <http://dx.doi.org/10.1016/j.nefro.2018.09.002>.
8. Zoccali C, Vanholder R, Massy ZA, Ortiz A, Sarafidis P, Dekker FW, et al. The systemic nature of CKD. *Nat Rev Nephrol*. 2017;13:344–58, <http://dx.doi.org/10.1038/nrneph.2017.52>.
9. Kannaiyan R, Mahadevan D. A comprehensive review of protein kinase inhibitors for cancer therapy. *Expert Rev Anticancer Ther*. 2018;1–22, <http://dx.doi.org/10.1080/14737140.2018.1527688>.
10. Ortiz A, Husi H, Gonzalez-Lafuente L, Valiño-Rivas L, Fresno M, Sanz AB, et al. Mitogen-activated protein kinase 14 promotes AKI. *J Am Soc Nephrol*. 2016, <http://dx.doi.org/10.1681/ASN.2015080898>.
11. Chak H, Ortiz A, Sanchez-Niño MD. MAGE genes in the kidney: identification of MAGE2 as upregulated during kidney injury and in stressed tubular cells. *Nephrol Dial Transplant*. 2018 Dec 11, <http://dx.doi.org/10.1093/ndt/gfy367> [Epub ahead of print] PubMed PMID: 30541139.
12. Matsuzawa A, Ichijo H. Redox control of cell fate by MAP kinase: physiological roles of ASK1-MAP kinase pathway in stress signaling. *Biochim Biophys Acta*. 2008;1780:1325–36, <http://dx.doi.org/10.1016/j.bbagen.2007.12.011>.
13. Gerits N, Kostenko S, Moens U. In vivo functions of mitogen-activated protein kinases: conclusions from knock-in and knock-out mice. *Transgenic Res*. 2007;16:281–314, <http://dx.doi.org/10.1007/s11248-006-9052-0>.
14. Chang L, Karin M. Mammalian MAP kinase signalling cascades. *Nature*. 2001;410:37–40, <http://dx.doi.org/10.1038/35065000>.
15. Sanz AB, Sanchez-Niño MD, Izquierdo MC, Jakubowski A, Justo P, Blanco-Colio LM, et al. Tweak induces proliferation in renal tubular epithelium: a role in uninephrectomy induced renal hyperplasia. *J Cell Mol Med*. 2009;13:3329–42, <http://dx.doi.org/10.1111/j.1582-4934.2009.00766.x>.
16. Zhuang Y, Feng Q, Ding G, Zhao M, Che R, Bai M, et al. Activation of ERK1/2 by NADPH oxidase-originated reactive oxygen species mediates uric acid-induced mesangial cell proliferation. *Am J Physiol Renal Physiol*. 2014;307:F396–406, <http://dx.doi.org/10.1152/ajprenal.00565.2013>.
17. Rayego-Mateos S, Morgado-Pascual JL, Sanz AB, Ramos AM, Eguchi S, Batlle D, et al. TWEAK transactivation of the epidermal growth factor receptor mediates renal inflammation. *J Pathol*. 2013;231:480–94, <http://dx.doi.org/10.1002/path.4250>.
18. Cargnello M, Roux PP. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol Mol Biol Rev*. 2011;75:50–83, <http://dx.doi.org/10.1128/MMBR.00031-10>.
19. Grynberg K, Ma FY, Nikolic-Paterson DJ. The JNK signaling pathway in renal fibrosis. *Front Physiol*. 2017;8:829, <http://dx.doi.org/10.3389/fphys.2017.00829>.
20. González-Guerrero C, Ocaña-Salceda C, Berzal S, Carrasco S, Fernández-Fernández B, Cannata-Ortiz P, et al. Calcineurin inhibitors recruit protein kinases JAK2 and JNK TLR signaling and the UPR to activate NF- κ B-mediated inflammatory responses in kidney tubular cells. *Toxicol Appl Pharmacol*. 2013;272:825–41, <http://dx.doi.org/10.1016/j.taap.2013.08.011>.
21. González-Guerrero C, Cannata-Ortiz P, Guerri C, Egidio J, Ortiz A, Ramos AM. TLR4-mediated inflammation is a key pathogenic event leading to kidney damage and fibrosis in cyclosporine nephrotoxicity. *Arch Toxicol*. 2017;91:1925–39, <http://dx.doi.org/10.1007/s00204-016-1830-8>.
22. Cardoso VG, Gonçalves GL, Costa-Pessoa JM, Thieme K, Lins BB, Casare FAM, et al. Angiotensin II-induced podocyte apoptosis is mediated by endoplasmic reticulum stress/PKC- δ /p38 MAPK pathway activation and through increased Na⁺/H⁺ exchanger isoform 1 activity. *BMC Nephrol*. 2018;19:179, <http://dx.doi.org/10.1186/s12882-018-0968-4>.
23. Liles JT, Corkey BK, Notte GT, Budas GR, Lansdon EB, Hinojosa-Kirschenbaum F, et al. ASK1 contributes to fibrosis and dysfunction in models of kidney disease. *J Clin Invest*. 2018;128:4485–500, <http://dx.doi.org/10.1172/JCI99768>.
24. Sanchez-Niño MD, Sanz AB, Ihalmo P, Lassila M, Holthofer H, Mezzano S, et al. The MIF receptor CD74 in diabetic podocyte injury. *J Am Soc Nephrol*. 2009;20:353–62, <http://dx.doi.org/10.1681/ASN.2008020194>.
25. Bera A, Das F, Ghosh-Choudhury N, Li X, Pal S, Gorin Y, et al. A positive feedback loop involving Erk5 and Akt turns on mesangial cell proliferation in response to PDGF. *Am J Physiol Cell Physiol*. 2014;306:C1089–100, <http://dx.doi.org/10.1152/ajpcell.00387.2013>.
26. Badshah II, Baines DL, Dockrell ME. Erk5 is a mediator to TGF β 1-induced loss of phenotype and function in human podocytes. *Front Pharmacol*. 2014;5:71, <http://dx.doi.org/10.3389/fphar.2014.00071>.
27. Déleris P, Trost M, Topisirovic I, Tanguay P-L, Borden KLB, Thibault P, et al. Activation loop phosphorylation of ERK3/ERK4 by group I p21-activated kinases (PAKs) defines a novel PAK-ERK3/4-MAPK-activated protein kinase 5 signaling pathway. *J Biol Chem*. 2011;286:6470–8, <http://dx.doi.org/10.1074/jbc.M110.181529>.
28. Wu D-D, Lau ATY, Yu F-Y, Cai N-L, Dai L-J, Ok Kim M, et al. Extracellular signal-regulated kinase 8-mediated NF- κ B activation increases sensitivity of human lung cancer cells to arsenic trioxide. *Oncotarget*. 2017;8:49144–55, <http://dx.doi.org/10.18632/oncotarget.17100>.
29. Malinin NL, Boldin MP, Kovalenko AV, Wallach D. MAP3K-related kinase involved in NF-kappaB induction by TNF, CD95 and IL-1. *Nature*. 1997;385:540–4, <http://dx.doi.org/10.1038/385540a0>.
30. Sanz AB, Sanchez-Niño MD, Izquierdo MC, Jakubowski A, Justo P, Blanco-Colio LM, et al. TWEAK activates the non-canonical NFkB pathway in murine renal tubular cells: modulation of CCL21. *PLoS One*. 2010;5, <http://dx.doi.org/10.1371/journal.pone.0008955>.
31. Sun S-C. The noncanonical NF- κ B pathway. *Immunol Rev*. 2012;246:125–40, <http://dx.doi.org/10.1111/j.1600-065X.2011.01088.x>.
32. Senftleben U, Cao Y, Xiao G, Greten FR, Krähn G, Bonizzi G, et al. Activation by IKK α of a second, evolutionary conserved, NF-kappa B signaling pathway. *Science*. 2001;293:1495–9, <http://dx.doi.org/10.1126/science.1062677>.
33. Busino L, Millman SE, Pagano M. SCF-mediated degradation of p100 (NF- κ B2): mechanisms and relevance in multiple myeloma. *Sci Signal*. 2012;5:pt14, <http://dx.doi.org/10.1126/scisignal.2003408>.
34. Xiong Y, Torsoni AS, Wu F, Shen H, Liu Y, Zhong X, et al. Hepatic NF-kB-inducing kinase (NIK) suppresses mouse liver regeneration in acute and chronic liver diseases. *Elife*. 2018;7, <http://dx.doi.org/10.7554/eLife.34152>.
35. Jung J-U, Ravi S, Lee DW, McFadden K, Kamradt ML, Toussaint LG, et al. NIK/MAP3K14 regulates mitochondrial dynamics and trafficking to promote cell invasion. *Curr Biol*. 2016;26:3288–302, <http://dx.doi.org/10.1016/j.cub.2016.10.009>.
36. Sheng L, Zhou Y, Chen Z, Ren D, Cho KW, Jiang L, et al. NF- κ B-inducing kinase (NIK) promotes hyperglycemia and glucose intolerance in obesity by augmenting glucagon action. *Nat Med*. 2012;18:943–9, <http://dx.doi.org/10.1038/nm.2756>.

37. Boutaffala L, Bertrand MJM, Remouchamps C, Seleznik G, Reisinger F, Janas M, et al. NIK promotes tissue destruction independently of the alternative NF- κ B pathway through TNFR1/RIP1-induced apoptosis. *Cell Death Differ*. 2015;22:2020–33, <http://dx.doi.org/10.1038/cdd.2015.69>.
38. Sanz AB, Sanchez-Niño MD, Ramos AM, Moreno JA, Santamaria B, Ruiz-Ortega M, et al. NF-kappaB in renal inflammation. *J Am Soc Nephrol*. 2010;21:1254–62, <http://dx.doi.org/10.1681/ASN.2010020218>.
39. Fernandez-Fernandez B, Izquierdo MC, Valiño-Rivas L, Nastou D, Sanz AB, Ortiz A, et al. Albumin downregulates Klotho in tubular cells. *Nephrol Dial Transpl*. 2018, <http://dx.doi.org/10.1093/ndt/gfx376>.
40. Ruiz-Andres O, Sanchez-Niño MD, Moreno JA, Ruiz-Ortega M, Ramos AM, Sanz AB, et al. Downregulation of kidney protective factors by inflammation: role of transcription factors and epigenetic mechanisms. *Am J Physiol Renal Physiol*. 2016;311:F1329–40, <http://dx.doi.org/10.1152/ajprenal.00487.2016>.
41. Moreno JA, Izquierdo MC, Sanchez-Niño MD, Suárez-Alvarez B, Lopez-Larrea C, Jakubowski A, et al. The inflammatory cytokines TWEAK and TNF α reduce renal klotho expression through NF κ B. *J Am Soc Nephrol*. 2011;22:1315–25, <http://dx.doi.org/10.1681/ASN.2010101073>.
42. Lorz C, Benito-Martín A, Boucherot A, Ucero AC, Rastaldi MP, Henger A, et al. The death ligand TRAIL in diabetic nephropathy. *J Am Soc Nephrol*. 2008;19:904–14, <http://dx.doi.org/10.1681/ASN.2007050581>.
43. Ruiz-Andres O, Suarez-Alvarez B, Sánchez-Ramos C, Monsalve M, Sanchez-Niño MD, Ruiz-Ortega M, et al. The inflammatory cytokine TWEAK decreases PGC-1 α expression and mitochondrial function in acute kidney injury. *Kidney Int*. 2016;89:399–410, <http://dx.doi.org/10.1038/ki.2015.332>.
44. González-Guerrero C, Morgado-Pascual JL, Cannata-Ortiz P, Ramos-Barron MA, Gómez-Alamillo C, Arias M, et al. CCL20 blockade increases the severity of nephrotoxic folic acid-induced acute kidney injury. *J Pathol*. 2018, <http://dx.doi.org/10.1002/path.5132>.
45. Poveda J, Sanz AB, Carrasco S, Ruiz-Ortega M, Cannata-Ortiz P, Sanchez-Niño MD, et al. Bcl3: a regulator of NF- κ B inducible by TWEAK in acute kidney injury with anti-inflammatory and antiapoptotic properties in tubular cells. *Exp Mol Med*. 2017;49:e352, <http://dx.doi.org/10.1038/emmm.2017.89>.
46. Poveda J, Sanz AB, Rayego-Mateos S, Ruiz-Ortega M, Carrasco S, Ortiz A, et al. NF κ Biz protein downregulation in acute kidney injury: modulation of inflammation and survival in tubular cells. *Biochim Biophys Acta*. 2016;1862:635–46, <http://dx.doi.org/10.1016/j.bbdis.2016.01.006>.
47. Sanz AB, Justo P, Sanchez-Niño MD, Blanco-Colio LM, Winkles JA, Kretzler M, et al. The cytokine TWEAK modulates renal tubulointerstitial inflammation. *J Am Soc Nephrol*. 2008;19:695–703, <http://dx.doi.org/10.1681/ASN.2007050577>.
48. Poveda J, Tabara LC, Fernandez-Fernandez B, Martin-Cleary C, Sanz AB, Selgas R, et al. TWEAK/Fn14 and non-canonical NF-kappaB signaling in kidney disease. *Front Immunol*. 2013;4:447, <http://dx.doi.org/10.3389/fimmu.2013.00447>.
49. Valiño-Rivas L, Gonzalez-Lafuente L, Sanz AB, Ruiz-Ortega M, Ortiz A, Sanchez-Niño MD. Non-canonical NF κ B activation promotes chemokine expression in podocytes. *Sci Rep*. 2016;6:28857, <http://dx.doi.org/10.1038/srep28857>.
50. Feng B, Chen G, Zheng X, Sun H, Zhang X, Zhang Z-X, et al. Small interfering RNA targeting RelB protects against renal ischemia-reperfusion injury. *Transplantation*. 2009;87:1283–9, <http://dx.doi.org/10.1097/TP.0b013e3181a1905>.
51. Benedetti G, Fokkelman M, Yan K, Fredriksson L, Herpers B, Meerman J, et al. The nuclear factor κ B family member RelB facilitates apoptosis of renal epithelial cells caused by cisplatin/tumor necrosis factor α synergy by suppressing an epithelial to mesenchymal transition-like phenotypic switch. *Mol Pharmacol*. 2013;84:128–38, <http://dx.doi.org/10.1124/mol.112.084053>.
52. Brightbill HD, Suto E, Blaquiére N, Ramamoorthi N, Sujatha-Bhaskar S, Gogol EB, et al. NF- κ B inducing kinase is a therapeutic target for systemic lupus erythematosus. *Nat Commun*. 2018;9:179, <http://dx.doi.org/10.1038/s41467-017-02672-0>.
53. Sanchez-Niño MD, Poveda J, Sanz AB, Mezzano S, Carrasco S, Fernandez-Fernandez B, et al. Fn14 in podocytes and proteinuric kidney disease. *Biochim Biophys Acta – Mol Basis Dis*. 2013;1832:2232–43, <http://dx.doi.org/10.1016/j.bbdis.2013.08.010>.
54. Martín P, Mora I, Cortes M, Calleros L, García-Jerez A, Ortiz A, et al. Relevant role of PKG in the progression of fibrosis induced by TNF-like weak inducer of apoptosis. *Am J Physiol Renal Physiol*. 2014;307:F75–85, <http://dx.doi.org/10.1152/ajprenal.00398.2013>.
55. Ucero AC, Benito-Martin A, Fuentes-Calvo I, Santamaria B, Blanco J, Lopez-Novoa JM, et al. TNF-related weak inducer of apoptosis (TWEAK) promotes kidney fibrosis and Ras-dependent proliferation of cultured renal fibroblast. *Biochim Biophys Acta*. 2013;1832:1744–55, <http://dx.doi.org/10.1016/j.bbdis.2013.05.032>.
56. Martínez-Miguel P, Medrano-Andrés D, Grier-Merino M, Ortiz A, Rodríguez-Puyol M, Rodríguez-Puyol D, et al. Tweak up-regulates endothelin-1 system in mouse and human endothelial cells. *Cardiovasc Res*. 2017;113:207–21, <http://dx.doi.org/10.1093/cvr/cvw239>.
57. Hénaut L, Sanz AB, Martin-Sanchez D, Carrasco S, Villa-Bellosta R, Aldamiz-Echevarria G, et al. TWEAK favors phosphate-induced calcification of vascular smooth muscle cells through canonical and non-canonical activation of NF κ B. *Cell Death Dis*. 2016;7:e2305, <http://dx.doi.org/10.1038/cddis.2016.220>.
58. Hénaut L, Sanchez-Niño MD, Aldamiz-Echevarría Castillo G, Sanz AB, Ortiz A. Targeting local vascular and systemic consequences of inflammation on vascular and cardiac valve calcification. *Expert Opin Ther Targets*. 2016;20:89–105, <http://dx.doi.org/10.1517/14728222.2015.1081685>.
59. Hoffmann A, Baltimore D. Circuitry of nuclear factor kappaB signaling. *Immunol Rev*. 2006;210:171–86, <http://dx.doi.org/10.1111/j.0105-2896.2006.00375.x>.
60. Hoesel B, Schmid JA. The complexity of NF- κ B signaling in inflammation and cancer. *Mol Cancer*. 2013;12:86, <http://dx.doi.org/10.1186/1476-4598-12-86>.
61. Bonizzi G, Beben M, Otero DC, Johnson-Vroom KE, Cao Y, Vu D, et al. Activation of IKK α target genes depends on recognition of specific kappaB binding sites by RelB:p52 dimers. *EMBO J*. 2004;23:4202–10, <http://dx.doi.org/10.1038/sj.emboj.7600391>.
62. Hoffmann A, Levchenko A, Scott ML, Baltimore D. The IkappaB-NF-kappaB signaling module: temporal control and selective gene activation. *Science*. 2002;298:1241–5, <http://dx.doi.org/10.1126/science.1071914>.
63. Tesch GH, Ma FY, Nikolic-Paterson DJ. ASK1: a new therapeutic target for kidney disease. *Am J Physiol Renal Physiol*. 2016;311, <http://dx.doi.org/10.1152/ajprenal.00208.2016>. F373-81.
64. Terada Y, Inoshita S, Kuwana H, Kobayashi T, Okado T, Ichijo H, et al. Important role of apoptosis signal-regulating kinase 1 in ischemic acute kidney injury. *Biochem Biophys Res Commun*. 2007;364:1043–9, <http://dx.doi.org/10.1016/j.bbrc.2007.10.122>.

65. El Eter E. NQDI 1, an inhibitor of ASK1 attenuates acute ischemic renal injury by modulating oxidative stress and cell death. *Cardiovasc Hematol Agents Med Chem*. 2013;11:179–86.
66. Ying W-Z, Wang P-X, Sanders PW. Pivotal role of apoptosis signal-regulating kinase 1 in monoclonal free light chain-mediated apoptosis. *Am J Pathol*. 2012;180:41–7, <http://dx.doi.org/10.1016/j.ajpath.2011.09.017>.
67. Ren G, Huynh C, Bijian K, Cybulsky AV. Role of apoptosis signal-regulating kinase 1 in complement-mediated glomerular epithelial cell injury. *Mol Immunol*. 2008;45:2236–46, <http://dx.doi.org/10.1016/j.molimm.2007.11.013>.
68. Amos LA, Ma FY, Tesch GH, Liles JT, Breckenridge DG, Nikolic-Paterson DJ, et al. ASK1 inhibitor treatment suppresses p38/JNK signalling with reduced kidney inflammation and fibrosis in rat crescentic glomerulonephritis. *J Cell Mol Med*. 2018;22:4522–33, <http://dx.doi.org/10.1111/jcmm.13705>.
69. Ucero AC, Benito-Martin A, Izquierdo MC, Sanchez-Niño MD, Sanz AB, Ramos AM, et al. Unilateral ureteral obstruction: beyond obstruction. *Int Urol Nephrol*. 2014;46:765–76, <http://dx.doi.org/10.1007/s11255-013-0520-1>.
70. Ma FY, Tesch GH, Nikolic-Paterson DJ. ASK1/p38 signaling in renal tubular epithelial cells promotes renal fibrosis in the mouse obstructed kidney. *Am J Physiol Renal Physiol*. 2014;307, <http://dx.doi.org/10.1152/ajprenal.00211.2014>. F1263–73.
71. Tesch GH, Ma FY, Han Y, Liles JT, Breckenridge DG, Nikolic-Paterson DJ. ASK1 inhibitor halts progression of diabetic nephropathy in Nos3-deficient mice. *Diabetes*. 2015;64:3903–13, <http://dx.doi.org/10.2337/db15-0384>.
72. Lin JH, Zhang JJ, Lin S-L, Chertow GM. Design of a phase 2 clinical trial of an ASK1 inhibitor GS-4997, in patients with diabetic kidney disease. *Nephron*. 2015;129:29–33, <http://dx.doi.org/10.1159/000369152>.
73. Perez-Gomez MV, Sanchez-Niño MD, Sanz AB, Zheng B, Martín-Cleary C, Ruiz-Ortega M, et al. Targeting inflammation in diabetic kidney disease: early clinical trials. *Expert Opin Invest Drugs*. 2016;1–14, <http://dx.doi.org/10.1080/13543784.2016.1196184>.
74. Loomba R, Lawitz E, Mantry PS, Jayakumar S, Caldwell SH, Arnold H, et al. The ASK1 inhibitor selonsertib in patients with nonalcoholic steatohepatitis: a randomized, phase 2 trial. *Hepatology*. 2017;67:549–59, <http://dx.doi.org/10.1002/hep.29514>.
75. <https://www.anzctr.org.au/Trial/Registration/TrialReview.aspx?id=375685>; accessed December 6, 2018 n.d.
76. Tarrant J, Billin A, Xiao Y. Serum soluble tumor necrosis factor receptor 1 (sTNFR1) associates with decline in estimated glomerular filtration rate (eGFR) slope in a phase 2 study of selonsertib in diabetic kidney disease (DKD). *J Am Soc Nephrol*. 2018;29:537.
77. Ma FY, Tesch GH, Ozols E, Xie M, Schneider MD, Nikolic-Paterson DJ. TGF- β 1-activated kinase-1 regulates inflammation and fibrosis in the obstructed kidney. *Am J Physiol Renal Physiol*. 2011;300:F1410–21, <http://dx.doi.org/10.1152/ajprenal.00018.2011>.
78. Zhou J, Zhong J, Huang Z, Liao M, Lin S, Chen J, et al. TAK1 mediates apoptosis via p38 involve in ischemia-induced renal fibrosis. *Artif Cells Nanomed Biotechnol*. 2018;1–10, <http://dx.doi.org/10.1080/21691401.2018.1442841>.
79. Zhou J, Fan Y, Zhong J, Huang Z, Huang T, Lin S, et al. TAK1 mediates excessive autophagy via p38 and ERK in cisplatin-induced acute kidney injury. *J Cell Mol Med*. 2018;22:2908–21, <http://dx.doi.org/10.1111/jcmm.13585>.
80. Karolak MJ, Guay JA, Oxburgh L. Inactivation of MAP3K7 in FOXD1-expressing cells results in loss of mesangial PDGFRB and juvenile kidney scarring. *Am J Physiol Renal Physiol*. 2018;315:F336–44, <http://dx.doi.org/10.1152/ajprenal.00493.2017>.
81. Yaomura T, Tsuboi N, Urahama Y, Hobo A, Sugimoto K, Miyoshi J, et al. Serine/threonine kinase Cot/Tpl2, regulates renal cell apoptosis in ischaemia/reperfusion injury. *Nephrology (Carlton)*. 2008;13:397–404, <http://dx.doi.org/10.1111/j.1440-1797.2008.00959.x>.
82. Naguro I, Umeda T, Kobayashi Y, Maruyama J, Hattori K, Shimizu Y, et al. ASK3 responds to osmotic stress and regulates blood pressure by suppressing WNK1-SPAK/OSR1 signaling in the kidney. *Nat Commun*. 2012;3:1285, <http://dx.doi.org/10.1038/ncomms2283>.
83. Perez-Gomez MV, Sanchez-Niño MD, Sanz AB, Martín-Cleary C, Ruiz-Ortega M, Egido J, et al. Horizon 2020 in diabetic kidney disease: the clinical trial pipeline for add-on therapies on top of renin angiotensin system blockade. *J Clin Med*. 2015;4:1325–47, <http://dx.doi.org/10.3390/jcm4061325>.
84. Fernandez-Fernandez B, Ortiz A, Gomez-Guerrero C, Egido J. Therapeutic approaches to diabetic nephropathy – beyond the RAS. *Nat Rev Nephrol*. 2014;10:325–46, <http://dx.doi.org/10.1038/nrneph.2014.74>.
85. Rojas-Rivera J, Ortiz A, Egido J. Antioxidants in kidney diseases: the impact of bardoxolone methyl. *Int J Nephrol*. 2012;2012:321714, <http://dx.doi.org/10.1155/2012/321714>.
86. Pillai P, Surenya RS, Nair SV, Lakshmanan V-K. Cancer kinases and its novel inhibitors: past present and future challenges. *Curr Drug Targets*. 2015;16:1233–45.
87. Talpaz M, Shah NP, Kantarjian H, Donato N, Nicoll J, Paquette R, et al. Dasatinib in imatinib-resistant Philadelphia chromosome-positive leukemias. *N Engl J Med*. 2006;354:2531–41, <http://dx.doi.org/10.1056/NEJMoa055229>.
88. Miura M. Therapeutic drug monitoring of imatinib, nilotinib, and dasatinib for patients with chronic myeloid leukemia. *Biol Pharm Bull*. 2015;38:645–54, <http://dx.doi.org/10.1248/bpb.b15-00103>.
89. Shamroe CL, Comeau JM. Ponatinib: a new tyrosine kinase inhibitor for the treatment of chronic myeloid leukemia and Philadelphia chromosome-positive acute lymphoblastic leukemia. *Ann Pharmacother*. 2013;47:1540–6, <http://dx.doi.org/10.1177/1060028013501144>.
90. Song C, Chai W, Song L, Yang Y, Lin J. Chloridobis(1,10-phenanthroline)zinc(II) tetra-chlorido(1,10-phenanthroline)bis-muthate(III) monohydrate. *Acta Crystallogr Sect E Struct Rep Online*. 2010;67:m109–10, <http://dx.doi.org/10.1107/S1600536810052682>.
91. Clark JD, Flanagan ME, Telliez J-B. Discovery and development of Janus kinase (JAK) inhibitors for inflammatory diseases. *J Med Chem*. 2014;57:5023–38, <http://dx.doi.org/10.1021/jm401490p>.
92. Burotto M, Manasanch EE, Wilkerson J, Fojo T. Gefitinib and erlotinib in metastatic non-small cell lung cancer: a meta-analysis of toxicity and efficacy of randomized clinical trials. *Oncologist*. 2015;20:400–10, <http://dx.doi.org/10.1634/theoncologist.2014-0154>.
93. de Azambuja E, Holmes AP, Piccart-Gebhart M, Holmes E, Di Cosimo S, Swaby RF, et al. Lapatinib with trastuzumab for HER2-positive early breast cancer (NeoALTTO): survival outcomes of a randomised, open-label, multicentre, phase 3 trial and their association with pathological complete response. *Lancet Oncol*. 2014;15:1137–46, [http://dx.doi.org/10.1016/S1470-2045\(14\)70320-1](http://dx.doi.org/10.1016/S1470-2045(14)70320-1).
94. Fallahi P, Di Bari F, Ferrari SM, Spisni R, Materazzi G, Miccoli P, et al. Selective use of vandetanib in the treatment of thyroid cancer. *Drug Des Devel Ther*. 2015;9:3459–70, <http://dx.doi.org/10.2147/DDDT.S72495>.

95. Genovese MC, Kremer JM, Kartman CE, Schlichting DE, Xie L, Carmack T, et al. Response to baricitinib based on prior biologic use in patients with refractory rheumatoid arthritis. *Rheumatology (Oxford)*. 2018;57:900–8, <http://dx.doi.org/10.1093/rheumatology/kex489>.
96. Wu P, Nielsen TE, Clausen MH. FDA-approved small-molecule kinase inhibitors. *Trends Pharmacol Sci*. 2015;36:422–39, <http://dx.doi.org/10.1016/j.tips.2015.04.005>.
97. Tuttle KR, Brosius FC, Adler SG, Kretzler M, Mehta RL, Tumlin JA, et al. JAK1/JAK2 inhibition by baricitinib in diabetic kidney disease: results from a Phase 2 randomized controlled clinical trial. *Nephrol Dial Transpl*. 2018;33:1950–9, <http://dx.doi.org/10.1093/ndt/gfx377>.
98. Gurevich F, Perazella MA. Renal effects of anti-angiogenesis therapy: update for the internist. *Am J Med*. 2009;122:322–8, <http://dx.doi.org/10.1016/j.amjmed.2008.11.025>.
99. Jhaveri KD, Wanchoo R, Sakhiya V, Ross DW, Fishbane S. Adverse renal effects of novel molecular oncologic targeted therapies: a narrative review. *Kidney Int Rep*. 2017;2:108–23, <http://dx.doi.org/10.1016/j.ekir.2016.09.055>.
100. Larkin J, Ascierto PA, Dréno B, Atkinson V, Liszkay G, Maio M, et al. Combined vemurafenib and cobimetinib in BRAF-mutated melanoma. *N Engl J Med*. 2014;371:1867–76, <http://dx.doi.org/10.1056/NEJMoa1408868>.
101. Uthuriague C, Thellier S, Ribes D, Rostaing L, Paul C, Meyer N. Vemurafenib significantly decreases glomerular filtration rate. *J Eur Acad Dermatol Venereol*. 2014;28:978–9, <http://dx.doi.org/10.1111/jdv.12322>.
102. Wanchoo R, Jhaveri KD, Deray G, Launay-Vacher V. Renal effects of BRAF inhibitors: a systematic review by the Cancer and the Kidney International Network. *Clin Kidney J*. 2016;9:245–51, <http://dx.doi.org/10.1093/ckj/sfv149>.
103. Teuma C, Perier-Muzet M, Pelletier S, Nouvier M, Amini-Adl M, Dijoud F, et al. New insights into renal toxicity of the B-RAF inhibitor, vemurafenib, in patients with metastatic melanoma. *Cancer Chemother Pharmacol*. 2016;78:419–26, <http://dx.doi.org/10.1007/s00280-016-3086-7>.
104. Jhaveri KD, Sakhiya V, Fishbane S. Nephrotoxicity of the BRAF inhibitors vemurafenib and dabrafenib. *JAMA Oncol*. 2015;1:1133–4, <http://dx.doi.org/10.1001/jamaoncol.2015.1713>.
105. Teuma C, Pelletier S, Amini-Adl M, Perier-Muzet M, Maucourt-Boulch D, Thomas L, et al. Adjunction of a MEK inhibitor to vemurafenib in the treatment of metastatic melanoma results in a 60% reduction of acute kidney injury. *Cancer Chemother Pharmacol*. 2017;79:1043–9, <http://dx.doi.org/10.1007/s00280-017-3300-2>.
106. Perico L, Mandalà M, Schieppati A, Carrara C, Rizzo P, Conti S, et al. BRAF signaling pathway inhibition podocyte injury, and nephrotic syndrome. *Am J Kidney Dis*. 2017;70:145–50, <http://dx.doi.org/10.1053/j.ajkd.2016.12.013>.
107. Maanaoui M, Saint-Jacques C, Gnemmi V, Frimat M, Lionet A, Hazzan M, et al. Glomerulonephritis and granulomatous vasculitis in kidney as a complication of the use of BRAF and MEK inhibitors in the treatment of metastatic melanoma: a case report. *Medicine (Baltimore)*. 2017;96:e7196, <http://dx.doi.org/10.1097/MD.00000000000007196>.
108. Woodhouse K, Farrow PR, Wilkinson R. Reversible renal failure during treatment with captopril. *Br Med J*. 1979;2:1146–7.
109. Collste P, Haglund K, Lundgren G, Magnusson G, Ostman J. Reversible renal failure during treatment with captopril. *Br Med J*. 1979;2:612–3.
110. Farrow PR, Wilkinson R. Reversible renal failure during treatment with captopril. *Br Med J*. 1979;1:1680.
111. Hoorntje SJ, Weening JJ, Kallenberg CG, Prins EJ, Donker AJ. Serum-sickness-like syndrome with membranous glomerulopathy in patient on captopril. *Lancet (London, England)*. 1979;2:1297.
112. Islam MR, Jimenez T, Pelham C, Rodova M, Puri S, Magenheimer BS, et al. MAP/ERK kinase kinase 1 (MEKK1) mediates transcriptional repression by interacting with polycystic kidney disease-1 (PKD1) promoter-bound p53 tumor suppressor protein. *J Biol Chem*. 2010;285:38818–31, <http://dx.doi.org/10.1074/jbc.M110.145284>.
113. Liang R, Shen XL, Zhang B, Li Y, Xu W, Zhao C, et al. Apoptosis signal-regulating kinase 1 promotes Ochratoxin A-induced renal cytotoxicity. *Sci Rep*. 2015;5:8078, <http://dx.doi.org/10.1038/srep08078>.